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Permeability Studies on the Red Blood Cell of the Ground-Hog, *Marmota monax*¹

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The high degree of permeability exhibited by red blood cells of the ground-hog, *Marmota monax*, to various sugars and polyhydric alcohols has been demonstrated by Parpart ('36) with the hemolysis method. These red cells are more permeable to sugars and polyhydric alcohols than human red cells which are not permeable to the 6-carbon polyhydric alcohols, D-Mannitol and D-Sorbitol. Since Faust ('60) has shown with the photoelectric method on human red cells that variations in pH and temperature greatly affect the rates of penetration of these substances, it had occurred to us to investigate this phenomenon with ground-hog red cells. If the rates of penetration of sugars and polyhydric alcohols into ground-hog red cells are affected in a manner similar to that of human red cells, then this would indicate that perhaps their mechanism of penetration is similar.

METHODS

Approximately 5 ml of blood was obtained by heart puncture from an ether-anesthetized ground-hog. This blood was then quickly placed into a flask containing 10 ml of 1% NaCl-PO₄ at pH 7. No anticoagulants were used. The blood suspension was then washed three times in 1% NaCl-PO₄ at pH 7. Aliquots of this suspension, which was placed in the refrigerator, were prepared for experimentation by the method previously described by Faust ('60). Although Bang and Ørskov ('31), Meldahl and Ørskov ('40), Mawe ('56) and Faust ('60) have observed that glucose penetrates into human red cells more slowly when the blood is not fresh, this did not seem to be the case with ground-hog red cells. There was no noticeable change in the permeability of these cells to sugars and polyhydric alcohols even

after they had been stored in the refrigerator for two days. However, washed ground-hog red cells that were more than two days old were never used for experiments.

The relative rates of penetration of various monosaccharides and polyhydric alcohols into ground-hog red cells were studied by means of a photoelectric densimeter, Parpart ('35). Isosmotic concentrations (0.3 M) of these compounds were prepared in an isotonic NaCl-phosphate buffer which was adjusted to a desired pH and then allowed to mutarotate. Exactly 40 mm³ of a washed red cell suspension was added to 6 ml of the substrate solution which was in the densimeter.

RESULTS

Figure 1 illustrates the effect of pH at 37°C on the rates of penetration to a diffusion equilibrium of some sugars and polyhydric alcohols into ground-hog red cells. The influence of pH on the rates of penetration of the monosaccharides and the polyhydric alcohols is similar. It is obvious that these compounds penetrate more slowly as the pH of their medium becomes more acid. This pH effect is reversible. Incidentally, a similar pH effect was observed on the extremely slow (approximately 40 times slower than D-glucose) rate of penetration of the amino acid, glycine.

Table 1 shows the temperature coefficients (Q_{10}) which were calculated from the rates of penetration of various sugars and polyhydric alcohols at 37°C and 27°C and at pH 7. The temperature coefficients

¹ This work was done at the Marine Biological Laboratory, Woods Hole, Mass. and supported by the Whitehall Foundation.

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for the sugars ranged between 2.0 and 2.8, whereas those for the polyhydric alcohols were between 1.2 and 1.6. Also from table 1 it can be seen that the rates of penetration of the polyhydric alcohols are approximately related in a linear manner to the number of carbon atoms in their molecule through 5 carbons. This

is not true for the various sugars that have been tested.

Neither the disaccharide sucrose nor the cyclic 6 carbon polyhydric alcohol i-inositol were observed to penetrate into the groundhog red cell. Also, there was no appreciable difference in the rates of penetration of the α - and β -isomers of D-glucose at pH 7 and 37°C.

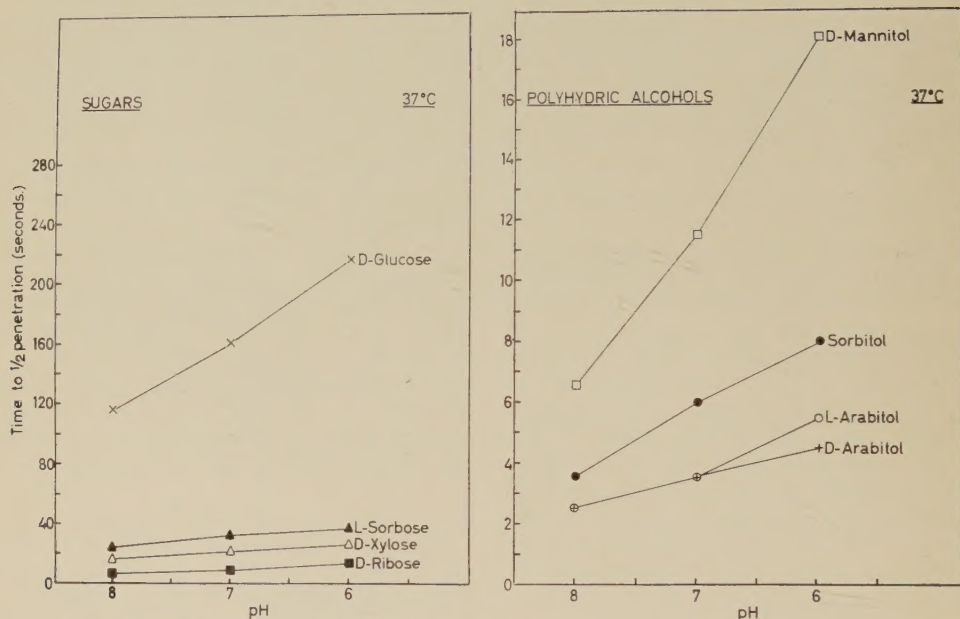


Fig. 1 Effect of pH on the relative rates of penetration of various 0.3 M sugars and polyhydric alcohols in 1% NaCl-PO₄. (Points represent an average of at least three determinations.)

TABLE 1

Results of Q_{10} studies on sugar and polyhydric alcohol penetration at pH 7

(0.3 M of substrate was dissolved in 1% NaCl-PO₄; sugars and alcohols are in order of decreasing penetration)

Sugar	No. of carbon atoms	$\frac{1}{2}$ Time penetration at		Q_{10}	Polyhydric alcohol	No. of carbon atoms	$\frac{1}{2}$ Time penetration at		Q_{10}
		37°C	27°C				37°C	27°C	
		seconds					seconds		
D-Ribose	5	9.0	21.0	2.3	Glycerol	3	1.0	1.5	1.5
D-Lyxose	5	16.0	36.0	2.2	i-Erythritol	4	2.1	3.0	1.4
D-Xylose	5	22.0	52.0	2.4	D-Arabitol	5	3.5	4.5	1.3
L-Xylose	5	24.0	48.0	2.0	L-Arabitol	5	3.5	4.5	1.3
D-Arabinose	5	24.0	52.0	2.2	D-Xylitol	5	3.5	4.5	1.3
L-Sorbose	6	33.0	72.0	2.2	Ribitol	5	4.0	5.0	1.3
D-Galactose	6	57.0	148.0	2.6	Sorbitol	6	6.0	8.0	1.3
D-Fructose	6	125.0	300.0	2.4	Galactitol	6	9.0	¹	
D-Glucose	6	160.0	450.0	2.8	D-Mannitol	6	11.5	16.0	1.4

¹ Galactitol was not sufficiently soluble at this temperature and could not be tested.

CONCLUSIONS

The effect of pH on the rates of penetration of 0.3 M sugars and polyhydric alcohols into ground-hog red cells is not the same as that observed by Faust ('60) with human red cells. These compounds penetrate into ground-hog red cells more rapidly as the pH of their medium becomes more alkaline. Whereas in human red cells, D- and L-arabitol and various monosaccharides penetrate more rapidly at pH 7, but, at pH 6 and 8, their rates of penetration are considerably reduced. Also, the effect of pH on the rates of penetration of these compounds is much greater in human red cells than in ground-hog red cells.

The temperature coefficients calculated for sugar penetration into ground-hog red cells is similar to that observed by Faust ('60) with human red cells. However, there exists only a small, if any, relationship between the structural and stereochemical differences among the monosaccharides and their relative rates of penetration into the ground-hog red cell.

These data may indicate that sugars and polyhydric alcohols penetrate into the ground-hog red cell by free diffusion. However, the free diffusion of certain large penetrating molecules may be affected by

an interaction between the molecule and the channel in the cell membrane through which it is passing.

ACKNOWLEDGMENT

The authors wish to thank Mr. Earl Van Norman for supplying the ground-hogs used in these experiments.

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The Endogenous Tissue Respiration of the Arctic Ground Squirrel as Affected by Hibernation and Season¹

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The Arctic ground squirrel, a hibernating species that normally spends upwards of 8 months of the year in an inactive or hibernating state, undergoes marked seasonal changes in body size and metabolism. Thus, after emerging from the burrow in late spring, and for the next month or so thereafter, he is at his minimum weight. Following a period of somewhat slow weight gain during the middle summer months, he quite suddenly starts to put on weight in the form of fat during the first few weeks of fall. Thus, sudden fattening brings him up to his maximum weight for the year. It just precedes the entrance into the burrow for the inactive portion of his yearly life cycle and it is here that he again loses weight in a series of hibernation and arousal periods.

Throughout the yearly cycle, these animals undoubtedly undergo marked shifts in the patterns of tissue metabolism. For instance, in the fall, just prior to hibernation, large amounts of fat are being deposited. Consequently, there must be a favoring of those pathways of metabolism leading from dietary carbohydrate and protein, as well as lipids, toward lipid synthesis. Conversely, during the winter months when the animals are losing weight, pathways of lipid catabolism must be favored. In addition, during the winter periods of alternating hibernation and arousal a number of peculiar, but for the most part unknown, processes of intermediary metabolism probably exist.

These obvious conclusions, however, are based almost entirely on deductive evidence. Direct investigations in the form of tissue respiratory measurements or enzyme activity assays are very limited in number. In fact, as far as could be determined, kid-

ney is the only tissue that has been studied with respect to its seasonal variations in metabolism (Kayser, Jacob and Lucot, '54).

It was felt, therefore, that a survey of the possible seasonal variations in the endogenous respiration of several tissues would be highly desirable. Such a survey would provide us with the background information and guidelines necessary to the design of future studies on isolated enzymes and enzyme systems.

METHODS

The Arctic ground squirrels (*Citellus undulatus*) used in this study were caught in the Little Susitna Valley of south central Alaska. While in captivity they were housed in individual wire cages and fed an ad libitum diet of whole kernel corn plus occasional lettuce and carrots. Water was available at all times. To induce hibernation, the animals were placed in a cold room ($10 \pm 1^\circ\text{C}$). At least 45 days were allowed to elapse before any hibernating or nonhibernating individuals were removed for study. As would be expected, the squirrels lost considerable weight during the course of the fall, winter, and spring studies. Thus, the weight range of individuals used in the fall was 628 to 825 gm whereas in the late winter or early spring the weight range was 305 to 430 gm. Five categories of individuals were selected: (1) before hibernation in the fall of the year; (2) one to three days hibernation; (3) 6 to 9 days hibernation;

¹ The views expressed are those of the authors and do not necessarily represent official Air Force policy. The experiments were conducted according to the "Rules Regarding Animal Care" as established by the American Medical Association (Air Force Regulation 160-94).

(4) one to two days after arousal from hibernation and (5) after completion of the hibernation season in the spring of the year. These animals were not segregated according to sex. However, no gross differences attributable to sex were observed in any of the measurements reported here.

The rats used for comparative purposes in these experiments were large (375 to 530 gm) adult males of the Sprague-Dawley strain. They were maintained in the animal colony at $26 \pm 1^\circ\text{C}$ on an ad libitum diet of "Friskies" dog food and water.

In these studies, endogenous oxygen consumption was measured using tissue homogenates rather than tissue slices, since it was felt that over a long period of time more consistent and comparable homogenate than slice preparations could be obtained. For comparative purposes, measurements on large albino rats were also made. Four different tissues were studied: the cerebrum lateral to the lateral ventricles, a portion of the large median lobe of the liver, the apex of the heart ventricles and a portion of the large muscle mass from the thigh of the hind leg. The following procedure was used to prepare the tissues for study: First, the animals were killed by decapitation and the desired tissues quickly excised and chilled in chopped ice. Aliquots of tissue were then taken and weighed on a microtorsion balance and homogenates prepared in ice-cold 0.25 M sucrose to give a final tissue concentration of 3%. The homogenates were filtered and 1-ml aliquots transferred to Warburg reaction vessels for oxygen consumption measurements.

The assay medium in the endogenous respiration studies had the following final concentration of constituents: 0.002 M K-ATP (pH 7.4), 0.01 M K-phosphate buffer (pH 7.4), 0.0066 M MgCl_2 ; 5×10^{-4} M diphosphopyridine nucleotide; 1.5×10^{-5} M cytochrome c and 0.20 M sucrose. Where citrate oxidation was studied in liver tissue, substrate was added to a final concentration of 0.01 M. The final volume of the reactant mixture including tissue was 3 ml. The reaction flasks also contained 0.2 ml of 5N NaOH plus a folded

piece of filter paper in the center well for the absorption of CO_2 .

Following a 10 minute period of thermal equilibration oxygen consumption measurements were conducted for 60 minutes at 38°C . The rate was calculated as microliters consumed per milligram of nitrogen per hour. Nitrogen was determined by nesslerizing an acid digested aliquot of the filtered homogenate.

Due to wide differences in subclass variances in the respiratory measurements, Fisher's t test could not be legitimately used in many instances for a statistical analysis of the data. Statistical evaluation was therefore made using the nonparametric, Mann-Whitney U test (Siegel, '56).

RESULTS

In the present experiments, the endogenous respiration of 4 tissues (brain, heart, liver and skeletal muscle) were investigated at 5 critical periods of the yearly life cycle of the Arctic ground squirrel. The results of these experiments are presented in table 1. Also included in the table are the average rates for each of the experimental groups. A statistical summary of these data is included in table 2.

Brain respiration. Cerebral tissue from the fattened, nonhibernating Arctic ground squirrel had about 60% as great a respiratory rate as similar tissue from the rat. In the squirrel the oxygen consumption of this tissue was not appreciably altered during the first few days of hibernation. After 6 to 9 days in the latter state, however, oxidative metabolism was depressed. Thus, levels of approximately one-half the prehibernation value were observed. It was further observed that arousal from hibernation produced little or no recovery of the respiratory rate that characterized the prehibernating state. To a minor extent such recovery of the initial rates did occur in the spring after the hibernation season was completed.

Heart respiration. The effects of season and hibernation on myocardial oxygen consumption were quite similar to those observed in brain tissue. Thus, short intervals of hibernation were again without appreciable effect, and longer periods of hibernation were associated with marked reduction in respiratory rate. With

TABLE 1

Tissue respiration¹ during various stages of the yearly life cycle of the Arctic ground squirrel

N	Squirrel hibernation cycle					
	Before hibernation	1-3 Days hibernation	6-9 Days hibernation	Arousal	After hibernation	Rat
Brain						
1	167	150	63	91	109	266
2	173	107	72	85	93	274
3	140	142	80	90	90	267
4	187	170		83	122	265
5	173	157			115	286
6	166	135				
Av.	168	144	72	87	106	272
Heart						
1	336	391	118	170	197	582
2	292	348	123	167	209	503
3	281	305	133	187	189	601
4	359	276		179	184	635
5	278	267			215	550
		279				
Av.	309	328	125	176	199	574
Muscle						
1	200	167	99	111	123	84
2	197	162	117	187	61	64
3	243	172	76	135	92	69
4	185	173	103	135	196	51
5	226	154				76
6	246					
Av.	216	166	99	142	118	69
Liver						
1	108	61	49	112	110	50
2	105	64	66	108	100	59
3	106	89	79	111	119	55
4	112	58	53	104	99	47
5	110	51			142	46
		46				
Av.	108	62	67	109	114	51
Liver citrate oxidation ²						
1	352	195	187	282		209
2	349	229	158	275		187
3	307	190	199	289		232
4	295	234	198	290		176
5	321	200				199
Av.	325	210	186	284		199

¹ Respiratory rates expressed in $\mu\text{l O}_2$ consumed/mg tissue N/hour.

² Corrected for endogenous respiration.

respect to the latter, it was observed that the myocardial respiratory rates after 6 to 9 days of hibernation were approximately one-third of those observed in the nonhibernating, or the 1-3 day hibernating state. At least partial restoration of the prehibernating oxygen consumption was again obtained during arousal periods and in the spring after completion of the hibernation season. A point of special in-

terest in these data on heart respiration, particularly as it might apply to the species variations in heart function at low temperature, was the markedly higher endogenous respiration exhibited by myocardial tissue from the rat as compared to squirrel. Thus, the rat exhibits almost twice as much respiratory activity as the prehibernating squirrel. Compared to the values obtained after long hibernation, after an

TABLE 2
Statistical summary¹ of tissue respiration data

Tissue	Hiber. cycle	1-3 days	6-9 days	Arousal	After	Rat
Brain (endogenous)	Before	n.s.	s	s	s	s
	1-3 days		s	s	s	
	6-9 days			n.s.	s	
	Arousal				n.s.	
Heart (endogenous)	Before	n.s.	s	s	s	s
	1-3 days		s	s	s	
	6-9 days			n.s.	s	
	Arousal				s	
Muscle (endogenous)	Before	s	s	s	s	s
	1-3 days		s	n.s.	n.s.	
	6-9 days			n.s.	n.s.	
	Arousal				n.s.	
Liver (endogenous)	Before	s	s	n.s.	n.s.	s
	1-3 days		n.s.	s	s	
	6-9 days			s	s	
	Arousal				n.s.	
Liver (citrate)	Before	s	s	s	—	s
	1-3 days		n.s.	s	—	
	6-9 days			s	—	

¹ Symbols used: n.s., not significant; s, significant, where $p \leq 0.05$ by the two-tailed Mann-Whitney U test (see Siegel, '56).

arousal from hibernation, or in the spring after the hibernation season is over, this species difference was even greater, i.e., three to 4 times higher rates in the rats than in the squirrels.

Muscle respiration. The seasonal and hibernation-associated changes in endogenous skeletal muscle respiration are also quite similar to those observed in the two preceding tissues. The only statistically significant difference to be noted in muscle was a 23% decrease in respiratory activity after the shortest interval of hibernation. As in the preceding tissues, however, the most marked reduction in oxygen consumption was obtained after longer intervals of hibernation. In the present instance, the rate of respiration after 6 to 9 days of hibernation is about one-half that observed before hibernation. Another basic difference between muscular tissue and brain and heart tissue was the comparative oxidative rates of squirrels and rats. Instead of the rat showing somewhat higher activities than the prehibernating squirrel just the opposite was observed; namely, rat muscle had about one-third the rate shown by squirrel muscle.

Liver respiration. In many ways, the responses of the liver tissue were quite

different from the other three tissues studied here. For instance, it was the only tissue where minimal endogenous respiratory rates were observed during the first few days of hibernation; there being no apparent tendency for a progressive decline as hibernation progressed. Liver was also the only tissue where there was a complete recovery of the prehibernation rates during periods of arousal or at the termination of the hibernation season. Like muscle, but unlike brain and myocardial tissue, the endogenous liver respiration of squirrels was considerably higher than that observed in the rat.

In addition to measuring the endogenous respiration, the capacity of the liver to oxidize citrate was also tested. As would be expected, this substrate markedly stimulated the respiratory rate of the tissues. In spite of the stimulation, however, the various qualitative changes observed previously in the endogenous measurements were also quite apparent when citrate was added. The only statistical difference between the two experimental procedures was the failure of the tissue from the aroused animal to fully recover the respiratory rate shown by the tissue from the

hibernating animal when citrate was sent.

DISCUSSION

In the foregoing data, numerous significant differences in endogenous tissue respiration have been observed. In general, these differences may be placed into three categories: (1) those that are attributable to season; (2) those that are attributable to hibernation; and (3) those that are attributable to species.

With regard to the first of these categories, it was observed in the nonhibernating squirrel that all tissues except liver inhibited their maximal respiratory rates the fall just prior to the hibernation season. These high rates declined over the course of the winter until minimal, or near minimal, rates were obtained in the spring. It is noteworthy that the intact hibernator has been reported (Kayser, '57) to exhibit a summer to winter decline in whole body metabolism that was similar to the tissue changes observed here. Kayser, Jacob and Lucot ('54), however, could not detect any comparable summer to winter change in the respiration of kidney tissue.

The present experiments give no information on mechanisms responsible for these decreases in tissue respiratory rates over the course of the winter. Such information will only be provided by more detailed studies of the enzymes and enzyme systems. Accordingly, it will be necessary to determine whether there is a decline in the concentration of certain enzymes, a decline in the concentration of readily oxidizable substrates, or a change in the nature of the substrates being oxidized at different times of the year.

The second category of effects observed here were those attributable to hibernation. Here it was shown that the hibernating state, sooner or later, led to a decline in the respiration of all the tissues studied. A similar decline in endogenous respiration has been reported for kidney tissue (Kayser, Jacob and Lucot, '54). It was not brought out by the present experiments that the magnitude of this effect is similar (about 40 to 60%) in all the tissues studied. Finally, it was shown that arousal, from hibernation was accompanied by at least a partial recovery of endogenous tissue respiration.

Again, it might be asked whether the alterations in tissue respiration can be attributed to changes in enzyme concentrations, to changes in substrate concentrations or to changes in the nature of the substrates being oxidized. In these experiments at least a partial answer to this question can be supplied. Thus, it was shown in liver tissue that the addition of citrate as substrate, although markedly stimulating respiration, did not alter the basic qualitative effects of hibernation on tissue metabolism. Since the citrate was added in excess amounts the observed rates would presumably reflect the catalytic capacity of the tricarboxylic acid cycle for its oxidation. It would follow, then, that the decline in liver oxidative metabolism upon entrance into the hibernating state was most likely due to a decline in the activity or concentration of one or more of the enzymes and cofactors that make up the tricarboxylic acid cycle and the electron transport system. Similarly, the recovery of the respiratory rate during the arousal process would be due to a recovery of the particular enzymes or cofactors that were affected by hibernation.

The foregoing results give no indication of the nature of the substrates being oxidized by the hibernating or aroused animal. Respiratory Quotient measurements on the intact animal, however, seem to indicate that the hibernating state is associated with lipid oxidation while the aroused state is associated with a return of carbohydrate oxidation (Dogden and Blood, '54; Mokrasch, Grady and Blood, '60). It will be of interest in future experiments to determine whether the loss of tissue oxidative capacity is directly attributable to the shift between carbohydrate and fat oxidation or to some other, as yet unknown, factors.

The final category of effects observed here were those attributable to species differences in tissue respiration. These differences included higher rates of brain and heart respiration and lower rates of liver and muscle respiration in the rat as compared to the nonhibernating squirrel. In these measurements it was interesting to observe that the lower rates of respiration are found in the two tissues that retain readily observable functional activity

during the hibernating state. In contrast to this, the tissues, especially muscle, that are least active during hibernation have the higher respiratory rates. This would seem to be something of a paradox since we would expect more active tissues to exhibit higher rates of respiration. In heart muscle, at least, these species differences are probably located at the enzyme level. Thus, it has been shown (Covino and Hannon, '59) that the ground squirrel myocardium had much less capacity to oxidize a variety of substrates than the rat myocardium at both normal body temperatures (38°C) and at hypothermic body temperatures (15°C). In fact, glutamate was the only substrate where the oxidative capacity of the squirrel myocardium exceeded that of the rat. How these hibernating animals are capable of maintaining heart and brain function at low temperatures in the face of this reduced capacity for energy release presents a most intriguing question. What makes it even more intriguing is the fact that these animals, in spite of their lower rate of myocardial respiration, are able to maintain not only higher concentrations of adenine nucleotides than the rat, but are also able to prevent the breakdown of ATP during hypothermia (Covino and Hannon, '59). There is also evidence (Zimny and Gregory, '59) that they can maintain reasonably high levels of myocardial ATP at least during the early stages of hibernation.

SUMMARY

The effects of season (autumn versus spring), short and long term hibernation, arousal from hibernation on the endogenous respiration of cerebral, myocardial, hepatic and skeletal muscle tissue of Arctic ground squirrels were studied. The species differences in tissue respiration between laboratory rats and ground squirrels were also investigated.

1. In the ground squirrels, all tissues except liver exhibited higher rates of endogenous respiration in the autumn than in the spring. Liver respiration was unaffected by season.

2. In all tissues, the hibernating state was associated with a reduction in endogenous respiration as measured at 38°C. This reduction was apparent after one to three days in muscle and liver tissue and after 6 to 9 days in brain and heart tissue. Arousal from hibernation produced a partial to complete recovery of the prehibernation respiratory rates. Experiments on the capacity of the liver to oxidize citrate suggested that these hibernation-induced changes in respiration were attributable to changes in the catalytic capacity of the tissue.

3. The brain and heart respiration was higher in the squirrel than the rat while the converse was true for liver and muscle tissue.

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Some Aspects of the Electoretinographic Response of the American Red Squirrel, *Tamiosciurus hudsonicus loquax*

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In the course of an investigation of the electoretinograms of various members of the family Sciuridae, some observations were made on the responses of the eye of the American red squirrel, *Tamiosciurus hudsonicus loquax*, which indicated that certain of its physiological reactions differ somewhat from those of other diurnal species of this family. It was also found that there are slight but apparently significant differences between the neural structure of the retina of the red squirrel and that of the ground squirrel.

It should be made clear at the outset that each of the diurnal species of the Sciuridae so far examined have been found to possess a pure-cone retina. This is true not only for the ground squirrels (*Citellus* spp.) of which 6 have been studied to date, but also for one chipmunk species (*Eutamias amoenus luteiventris*), for the prairie dog (*Cynomys ludovicianus*) and for three species of tree squirrel, the American red squirrel, *Tamiosciurus hudsonicus loquax*, the European red squirrel, *Sciurus vulgaris* and the common gray squirrel, *Sciurus hudsonicus leucotis*. The retinæ of the tree squirrels differ from those of the *Citellus* species, the chipmunk and the prairie dog in having two layers of cones instead of the more usual single layer, but otherwise the general structure of all these pure-cone retinæ is very similar, such differences as there are especially between the ground squirrels and the American red squirrel, being a matter of the relative numbers of cells to be found in the various clear layers. This probably means differences in the amounts of differentiation in Granit's ('47) sense) and summation

between the *Citellus* retina and that of the red squirrel. The electrophysiological results to be reported here suggest that such differences really do exist.

In this paper we are reporting on the dark adaptation and flicker responses of the red squirrel as measured by means of the electoretinogram. The results are considered in relation to those obtained in similar experiments on two ground squirrel species, *Citellus lateralis* and *Citellus beecheyi* and one chipmunk species, *Eutamias amoenus luteiventris*.

METHODS

The preparation of the animals, anesthesia, placement of the electrodes, light stimulus and general experimental details have already been described (Tansley, Copenhaver and Gunkel, '60). The Grass electroencephalograph was used for recording the responses during the course of dark adaptation, but this instrument was too slow accurately to follow the responses to flicker. For these experiments, therefore, the responses were amplified and displayed on an ordinary cathode ray oscilloscope and photographed on moving film. The time constant of the amplifier was 900 msec.

For the dark adaptation curves the eye was adapted for 5 minutes to the full stimulus (7500 lux) with the room lights on. The room lights and the adapting light were switched off simultaneously and records were taken at measured intervals of about 5 seconds each for three minutes, using the full stimulus with a 1.2 neutral density filter and a flash duration of 250 msec. The actual timing at which

each stimulus was given was read off afterwards on the paper. Curves were constructed relating the changes in the *a*-wave, *b*-wave and off-effect to the period of dark adaptation. All changes were complete in three minutes.

For the flicker experiments a sectorized disc was made giving equal duration light and dark intervals. The speed of the motor running this disc could be continuously varied to give stimulus frequencies varying between 17 and 170 flashes per second. A photocell placed between the disc and the eye was connected up so as to record on the second beam of the oscilloscope. The disc was then run at a speed slow enough to give an obviously intermittent response on the oscilloscope. The camera was then started and the speed of the disc increased until the response was quite steady. The actual speed at which fusion first took place was read off on the film after development. This procedure was repeated starting with the full stimulus intensity and continuing with increasingly dense neutral filters in the light beam until the stimulus was too small to evoke a response. The stimulus intensity was then increased by the same steps until its full intensity was again reached. The fusion frequency at each stimulus intensity was then read off from the film after development and the mean of the values for the two series "up" and "down" calculated. The crystal-line lens was not removed in these experiments which were done on the dark-adapted animal.

When recording was complete the upper part of the animal was intravitally fixed through the ascending aorta after ligaturing the descending aorta in the chest and washing out with isotonic saline. Bouin's fluid was used for fixation. The eyes were then removed and left overnight in the fixative. The eyes were embedded whole in celloidin without removal of the lens and sections cut at 15 μ ; they were stained with hematoxylin and eosin. All sections were cut at right angles to the elongated nerve head characteristic of the squirrel eye (Walls, '42) so that it was possible to choose comparable sections from the eyes of each species for comparison.

RESULTS

Dark adaptation curves. In figure 1 the changes in the *a*-wave, *b*-wave and off-effect of one of the ground squirrels, *Citellus beecheyi*, during the first 180 minutes of dark adaptation are shown. It will be seen that all three waves increase in amplitude during dark adaptation, though the change in the *a*-wave at stimulus intensity is very small. Figure 2 gives the same curves for the red squirrel. In this animal the off-effect decreases during dark adaptation; for the first few seconds it is actually bigger than the *b*-wave but subsequently it gets smaller as the *b*-wave increases and after about 10 seconds its amplitude falls below that of the *b*-wave. Some actual records from other experiments on the same two species are shown in figure 3. Here the records in

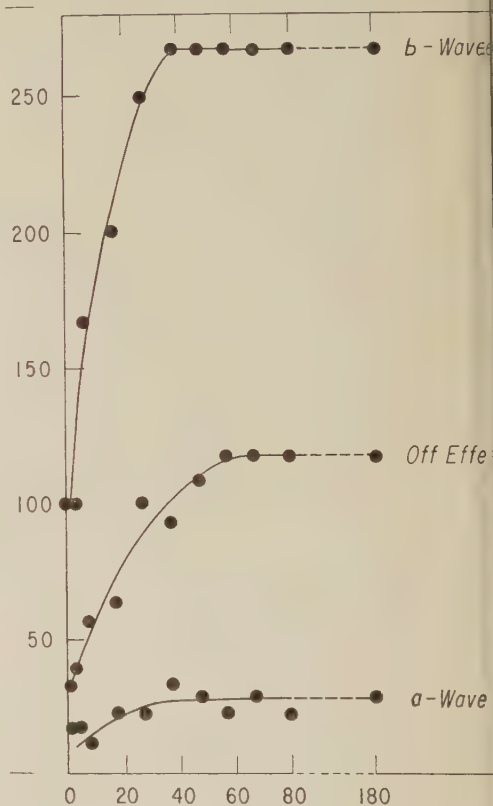


Fig. 1 Dark adaptation curves. The change in amplitude of the *a*-wave, *b*-wave and off-effect of the ground squirrel, *Citellus beecheyi*, during dark adaptation. Ordinates: potential development in μv . Abscissae: time in the dark in seconds.

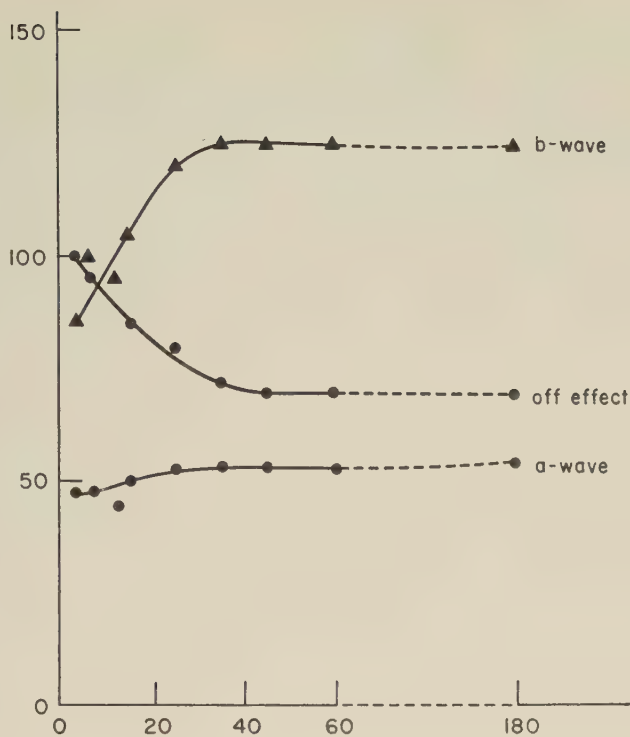


Fig. 2 Dark adaptation curves. The changes in amplitude of the a-wave, b-wave and off-effect of the American red squirrel, *Tamiosciurus hudsonicus loquax*, during dark adaptation. Ordinates: potential developed in μv . Abscissae: time in the dark in seconds.



Fig. 3 Changes during dark adaptation. Responses during the first 30 seconds of dark adaptation. A, American red squirrel, *Tamiosciurus hudsonicus loquax*. The arrows at two seconds and 5 seconds mark the b-wave. B, Ground squirrel, *Citellus beecheyi*; 0.25 second stimulus in both cases. The paper speed was rather faster in B.

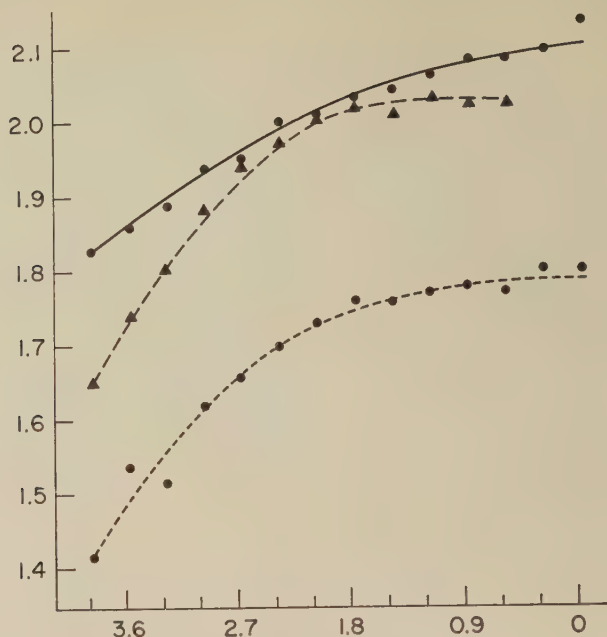


Fig. 4 Flicker fusion frequencies. The increase in fusion frequency with increased stimulus intensity. ●—● ground squirrel, *Citellus lateralis*; ▲---▲ chipmunk, *Eutamias amoenus luteiventris*; ●---● American red squirrel, *Tamiosciurus hudsonicus loquax*. Ordinates: log fusion frequency. Abscissae: neutral filter densities.

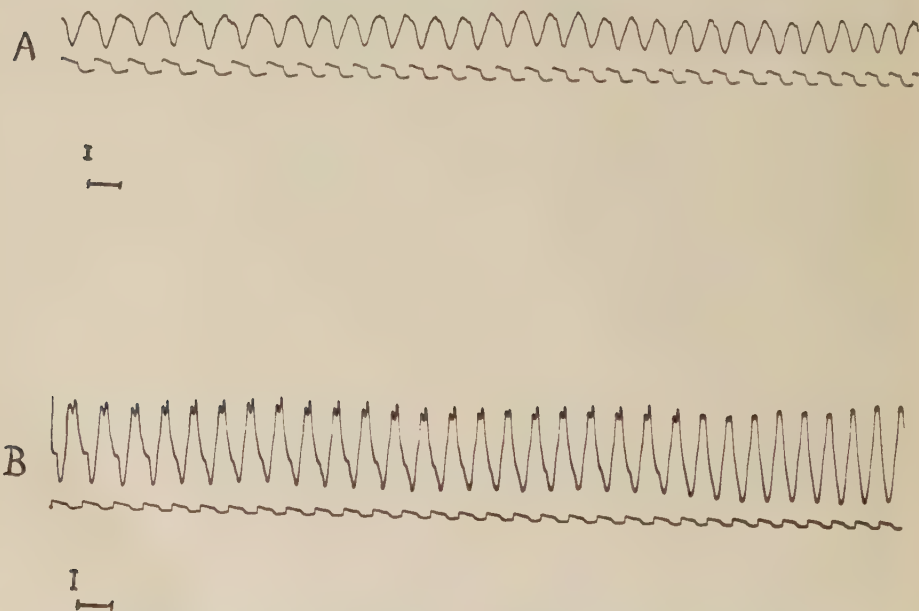


Fig. 5 Responses to flicker. A, American red squirrel, *Tamiosciurus hudsonicus Loquax*. B, Ground squirrel, *Citellus lateralis*. Calibration 100 μ v. Time marking 50 msec. Stimulus intensity 7500 lux.



Fig. 6 Section through the central retina of the ground squirrel, *Citellus beecheyi*. Bouin. Hematoxylin and eosin. $\times 315$. N.B. The following numbering applies to this and to figures 7 and 8. 1, pigment epithelium; 2, visual cells; 3, outer nuclear layer; 4, outer fiber layer; 5, inner nuclear layer; 6, inner fiber layer; 7, ganglion cell layer; 8, optic nerve fiber layer.

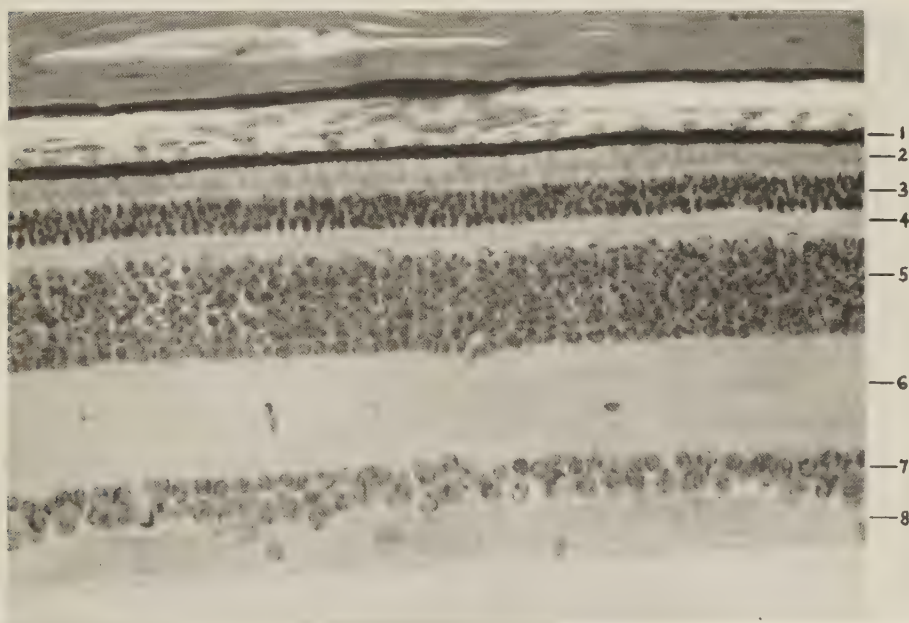


Fig. 7 Section through the central retina of the American red squirrel, *Tamiosciurus hudsonicus loquax*. Bouin. Hematoxylin and eosin. $\times 315$.

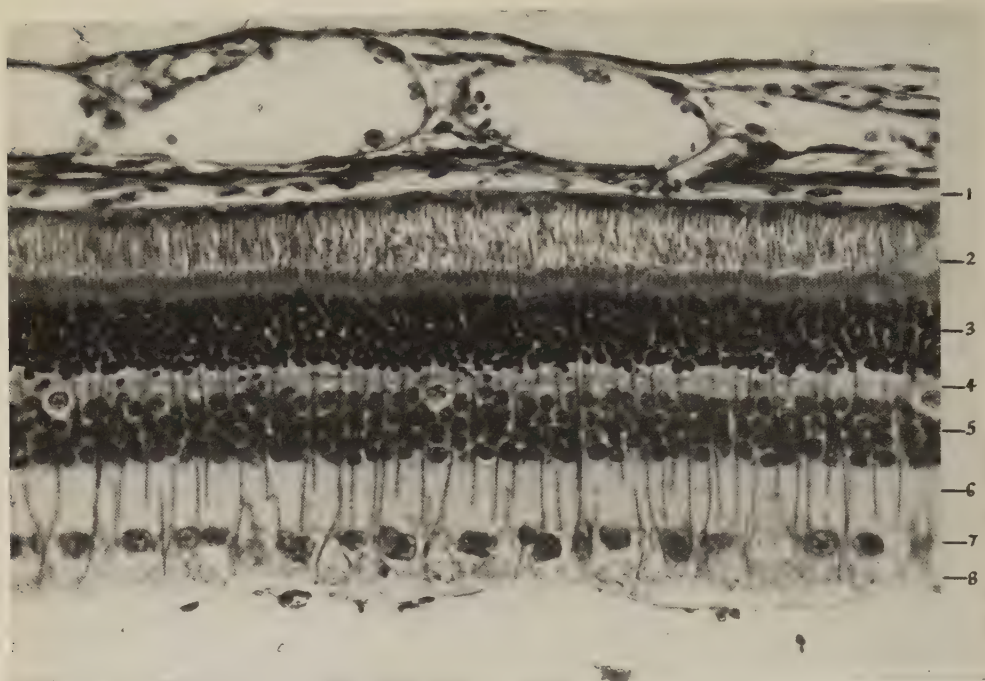


Fig. 8 Section through the central retina of the rabbit. Zenker. Mallory's phosphotungstic acid hematoxylin. $\times 294$.

are from the red squirrel, those in B from the ground squirrel. In the latter the off-effect always remains smaller than the *b*-wave, but in the red squirrel the off-effect begins by being most obviously the bigger, after about 15 seconds the two deflexions are approximately equal and by 30 seconds the off-effect is definitely smaller than the *b*-wave. This was a constant finding for the red squirrel under the conditions of our dark adaptation experiments, but we never saw the effect in any other squirrel species. In all these the off-effect remained smaller than the *b*-wave and increased in amplitude during dark adaptation. In all these pure-cone retinæ the changes due to dark adaptation were complete in 60 seconds or less. Such a rapid dark adaptation has already been reported for another pure-cone retina, that of the gray squirrel, *Sciurus carolinensis leucotis* (Tansley, '57).

Flicker. The curves in figure 4 show the increase in flicker fusion frequency with increased stimulus intensity for one specimen of the ground squirrel, *Citellus lateralis*, (top curve), a chipmunk (middle

curve) and a red squirrel (bottom curve). All the ground squirrels and the chipmunk investigated gave fusion frequencies of between 95 and 120 per second with our maximum intensity stimulus, but we never obtained a figure above 65 per second for the red squirrel under the same conditions. This difference is undoubtedly due to the fact that the red squirrel's responses to light are slower than those of the ground squirrels. With our maximum stimulus the implicit (peak) time of the *a*-wave was 15 msec and that of the *b*-wave 35 msec for the red squirrel. The figures for *Citellus beecheyi* were 10 and 15 msec respectively with the same stimulus. Some records of flicker responses for the red squirrel (A) and *Citellus lateralis* (B) are shown in figure 5. These records also show how much more precisely the ground squirrel retina can follow the stimulus changes.

Histology. Figure 6 shows a section through the highly developed central retina of a *Citellus beecheyi* retina. This area forms a strip lying under the elongated nerve head and stretching right across the fundus. The unusually thick inner nuclear

and ganglion cell layers are very noticeable. The section from a red squirrel shown in figure 7 was most carefully chosen from a comparable part of the retina. It is shown at the same magnification. It is plain that neither the inner nuclear layer nor the ganglion cell layer are so well developed in this retina. A section through the central area of the scotopic (rod-dominated) retina of a rabbit is shown in figure 8 for comparison.

DISCUSSION

Our curves relating flicker fusion frequency to stimulus intensity in figure 4 are very similar in shape to those published by Bornschein and Szegvari ('58) for another ground squirrel species, *Citellus citellus*. These authors used an electronic flash of very high intensity and a very short duration, which was independent of the flicker rate, for their stimulus and, in consequence, their intensity range is quite different from ours. However, their maximum fusion frequencies are very nearly identical with those which we recorded for the chipmunk and the *Citellus* species. Our red squirrel curve, although similar in shape, is at a generally lower frequency level. All these curves obtained from pure-cone retinæ are quite different from those reported by Dodt and Wirth ('53) and by Bornschein and Szegvari for the rod-dominated retina of the guinea pig. In this animal the curve is in two parts showing a scotopic maximum fusion frequency of about 20 per second and another at higher intensities when the fusion frequency can go up to about 50 per second. There is no sign of any such break either in our curves or in those shown by Bornschein and Szegvari.

As a result of his work on different types of retina, both rod-dominated and cone-dominated, Dodt has come to the conclusion that one can use the flicker fusion frequency as a means of differentiating between rod and cone responses. He believes that a fusion frequency above about 30 per second must be mediated by cones, and in a paper published in collaboration with Walther (Dodt and Walther, '58) uses the idea in the interpretation of some results obtained with the rabbit. The hypothesis may be correct although it is hard

to understand how the comparatively small number of cones present in the rabbit retina can completely dominate the electroretinographic response in the way postulated by Dodt. Even if one assumes that the cones can inhibit the rod response as suggested by Elenius and Heck ('57) to explain some of their results in man, there would still appear to be too few cones in the rabbit to give a measurable electroretinogram on their own. If one accepts Dodt's hypothesis there is quite a lot of variation among the cone responses in different species with regard to the fusion frequencies they can achieve. Dodt and Wirth report a fusion frequency of 140 per second for the far from pure-cone retina of the pigeon using a one-to-one light-dark ratio at 8000 lux. These conditions are very like ours since we also used a one-to-one light-dark ratio at 7500 lux, but we never recorded so high a figure. And with these same conditions the red squirrel figure never went above 65 per second. So although the value for this animal falls within Dodt's range for cones it is very markedly lower than that recorded for other cone-dominated retinæ under comparable conditions.

It is obvious that the relatively low fusion frequency of the red squirrel retina is a consequence of its slower responses to stimulation so that it is unable accurately to follow quick alternations of light and dark. But why should this particular pure-cone retina be slower in its reactions than the other pure-cone retinæ of the squirrel family? There is no reason to believe that this retina contains rods in sufficient number to affect the electroretinogram. Leaving aside for the moment a fusion frequency which Dodt would consider to be well outside the rod range, the criteria already discussed (Tansley, '57) in connection with the visual cells of the grey squirrel also apply to the red. This retina shows no Purkinje shift and has a rapid and not very extensive dark adaptation. In addition its spectral sensitivity curve is the same as that found in the grey squirrel and in the ground squirrels and chipmunk (Tansley, Copenhaver and Gunkel, '60).

It seems more likely that the answer lies in the neural organization of the retina.

If we compare a section of the red squirrel retina (fig. 7) with one from a ground squirrel (fig. 6) it is obvious that the latter shows the more complex structure. Not only are there enough ganglion cells to give a one-to-one relation between visual cells and optic nerve fibers but there are many more cells in the inner nuclear layer. Such an increase in complexity is typical of a cone-dominated retina in comparison with a rod-dominated one as can be seen from the section of a rabbit retina shown in figure 8. This section is also from the central retina where ganglion and bipolar cells are most numerous. In this rod retina it is clear that there can be no question of a one-to-one relationship between rods and optic nerve fibers and that, so far from there being more inner nuclear cells than there are visual cells as in the cone retina, there are in fact many fewer.

Granit ('55) has shown that inhibition is much better developed in the cone-dominated than in the rod-dominated retina and he argues that this is the reason for the much faster responses of the former. Since one of the most striking differences between the two types of retina is in the number of inner nuclear cells it seems probable that the increase in these seen in cone retinæ is associated with an increase in the amount of inhibition that can be developed. We do not know whether this increase in the number of cells in the inner nuclear layer is mainly due to an increased number of bipolar cells and, if so, of which types of bipolar. There is certainly some increase in horizontal cells and possibly also of amacrine cells. If we associate fast reactions with increased inhibition and this with an increased complexity of the inner nuclear layer then it would be natural to expect that the visual reactions of the red squirrel would be slower than those of the ground squirrel and that its flicker fusion frequency would be lower under comparable experimental conditions.

Another striking difference between the rod retina (fig. 8) and the cone retina (fig. 6) lies in the proportions of visual cells to ganglion cells. In the rod retina there are enormously more of the former and this must mean that a very large number of visual cells must be finally connected to

each ganglion cell and so to each optic nerve fiber of which the ganglion cell is the nucleus. Such an arrangement provides for a high degree of summation of visual cell responses onto each optic nerve fiber and this seems to be the basis of the great sensitivity of the rod retina as compared to the cone retina in which there can be nothing like so much summation. A comparison of the red squirrel retina (fig. 7) with that of the ground squirrel (fig. 6) indicates that there is likely to be more summation in the former and one would therefore, expect the red squirrel to be more sensitive to low illuminations. On the other hand the ground squirrel retina should be capable of greater resolution owing to its possession of more ganglion cells and therefore of more optic nerve fibers. We have no information as to the relative visual acuities of the two species but it does appear that the red squirrel has a higher sensitivity. Whereas the ground squirrels and chipmunks are reported only to be active during the hours of full daylight, we observed that the red squirrel does not at all confine its activity to the hours between sunrise and sunset. Three individuals kept in an outdoor cage regularly played on a revolving wheel in the evenings through the twilight until nightfall.

We are unable to explain why the off-effect in the red squirrel should be larger in the light-adapted state than during dark adaptation when in the ground squirrel the reverse is the case. It is true that in the frog Granit and Riddell ('34) long ago showed that the off-effect is much increased during light adaptation. They also found that the purely negative response produced by massage of the bulb and believed to be an expression of PIII activity, is increased by light adaptation and that it shows a quicker disappearance at "off" under these conditions. This was one of the findings considered by Granit when, in his well-known analysis of the electroretinogram, he suggested that the off-effect is largely due to the quicker disappearance of PIII than PII on cessation of illumination resulting in the transitory reappearance of a positive potential at "off." However, such an explanation cannot, as Granit and Therman ('37) first pointed

at and as has been shown much more recently by Tansley, Copenhaver and Gunkel (1961), account entirely for the large off-effect of the cone retina. There must, in addition, be a reactivation of PII at "off." It may be that such a reactivation of PII is a more important component of the ground squirrel off-effect while the interaction of PIII and PII plays a greater part in the red squirrel. Some such hypothesis might explain the difference in response to dark adaptation of the off-effect of the red squirrel as compared with that of the other squirrel species.

SUMMARY

1. Some electroretinographic responses of the American red squirrel were compared with those of two ground squirrel and one chipmunk species.
2. It was found that whereas the off-effect of the ground squirrel electroretinogram increased during dark adaptation that of the red squirrel decreased.
3. The flicker fusion frequency of the red squirrel as measured by the electroretinogram was consistently lower than that of the ground squirrels and chipmunk under the same experimental conditions.
4. The flicker results are discussed in the light of the retinal histology of the red and ground squirrels.
5. A tentative suggestion is put forward to explain the different behavior of the off-effect in light and dark adaptation as

between the red squirrel and the ground squirrels.

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Reversible and Irreversible Changes in Normal Human Thrombocyte Membranes by Slow and Rapid Swelling

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In previous communications (Gurevitch and Nelken, '56; Gurevitch, Nelken and Danon, '58) we described the morphological changes occurring in normal human blood platelets when introduced into solutions of decreasing ionic strength. The thrombocytes showed an increase in size; they became swollen and the spicules became longer and broader. In the solutions of very low ionic strength a peculiar thrombocyte shape appeared; the thrombocyte possessing a sword-like protrusion. Recently the osmotic fragility of normal human blood platelets has been investigated (Nelken, Gilboa-Garber and Gurevitch, '60). The morphological changes of the thrombocytes as well as the release of the enzyme pyrophosphatase into the hypotonic solutions were measured as a function of the decreased ionic strength. An analogy could be drawn between the classical osmotic fragility test of erythrocytes and the osmotic fragility of thrombocytes. In the osmotic fragility test of erythrocytes the cells are directly introduced into the hypotonic salt solution and the degree of hemolysis measured. When the hemolysis is performed through the gradual decrease in ionic strength (Danon, Nevo and Marikovsky, '56) and the degree of hemolysis measured (Klibansky, '60), the results obtained differ from those of the first "drastic" method. The ionic strength at which the hemoglobin is released through the red blood cell membrane in the dialysis procedure is lower than in the drastic or classical hemolysis method. The difference in the ionic strength at which the hemolysis occurs when erythrocytes of the same donor were examined by the drastic and gradual methods is attributed to

the visco-elastic nature of the erythrocyte membrane. The membrane is unable to resist the sudden pressure increase which occurs when the cell is introduced into a solution of low ionic strength. On the other hand, the red cell membrane withstands the slow increase of the internal pressure during the dialysis method down to a lower external salt concentration than in the drastic method.

In the present study an attempt has been made to examine normal human thrombocytes exposed to a gradual decrease in ionic strength as compared with thrombocytes exposed to "drastic" reductions in outer solution concentration.

MATERIALS AND METHODS

Preparation of thrombocyte suspension

The blood drawing, the anticoagulant solution and the method of separation of the thrombocytes have been described elsewhere (Nelken, Gilboa-Garber and Gurevitch, '60).

The decrease of ionic strength by the gradual method

Gradual decrease in the salt concentration surrounding the thrombocytes was achieved by dialysing a suspension of thrombocytes in physiological salt solution against the desired hypotonic solution in the following manner: Thrombocytes separated from 10 ml of blood were introduced into a dialysis bag¹ containing 10 ml of 0.15 N NaCl solution. The bag was suspended into a beaker containing 2000 ml

¹ Purchased from Fisher Scientific Company, New York, no. 8-667.

of 0.024 N NaCl solution. Another dialysis tube containing an equal volume of salt solution of the same concentration was suspended into the beaker at the same time. Every 5 to 10 minutes, 1 ml aliquot of the thrombocyte suspension was withdrawn and fixed in 1% (final concentration) osmium tetroxide in a NaCl solution of approximately the same ionic strength as the suspending medium of the aliquot, and buffered to pH 7.3 according to Palade '52. (Each batch of dialysis tubing was calibrated prior to performing thrombocyte experiments by establishing the decrease of ionic strength with time, under the experimental conditions stated.)

The "drastic" method

The "drastic" decrease in the salt concentration of the medium surrounding the thrombocytes was obtained by adding 0.05 ml aliquots of a concentrated stock suspension of thrombocytes in 0.15 N NaCl solution into 1 ml of NaCl solution taken from the second dialysis bag mentioned above (that containing no thrombocytes). As the solution in this dialysis bag may be presumed to have reached the same degree of hypotonicity as the one in the dialysis bag containing the thrombocytes undergoing gradual environmental changes, a direct comparison between the two methods under identical final conditions is possible. After 5 minutes in the hypotonic medium the thrombocytes were fixed by adding osmium tetroxide solution as described above.

After 20 minutes fixation, one drop of every suspension was examined between slide and cover glass under a phase-contrast microscope. One hundred cells were classified according to their different forms. The percentage of normal cells left in every sample as a function of ionic strength is shown in figure 1. The electron microscopical preparations were made by depositing a drop of suspension on a copper grid covered with a Formvar film. The drop was air dried, then washed with 10 successive drops of glass doubly-distilled water. After air drying the grids, they were shadow cast by platinum at a height to shadow ratio of 1 to 5. An RCA EMU 3B electron microscope was used.

Pyrophosphatase determination

The pyrophosphatase released from the thrombocytes into the surrounding NaCl solution was determined in 1-ml sample according to a method previously described (Nelken, Gilboa-Garber and Gurevitch, '60).

Reversion to isotonicity

Gradual reversion to isotonicity was achieved by placing the dialysis bag containing the remaining thrombocytes in isotonic strength solution into 1000 ml 0.1 N NaCl solution.

Rapid reversion was accomplished by adding hypertonic NaCl solution to each hypotonic salt solution containing the thrombocytes bringing it back to isotonicity.

Clot retraction experiments

The clot retraction was determined by adding 0.4 ml of the platelet suspension to 0.4 ml of own plasma, 0.05 ml of 30% suspension of thrice washed erythrocytes and 0.1 ml of 4.5% CaCl_2 . The clot retraction was examined after three hours and 24 hours. (The erythrocytes were introduced in order to facilitate the visualization of the degree of clot retraction.)

RESULTS

I. The morphological examination

Phase-contrast microscope observations

The morphological examination with the aid of the phase-contrast microscope showed no fundamental morphological difference between the platelets in the dialysis bag where hypotonicity was achieved gradually and the platelets introduced directly into the hypotonic solutions. When the NaCl concentration decreased to 0.1 N (after 10 minutes dialysis), an overall swelling of the platelets was observed. The same morphological changes, but more exaggerated, were seen in the platelets that were directly introduced into the hypotonic solution of the same ionic strength.

When the salt concentration reached 0.10 N (after 20 minutes of dialysis) the swelling was more pronounced.

When the NaCl solution went down to 0.083 N (30 minutes dialysis) all the platelets from the dialysis bag were still without spicules, and may thus still be considered morphologically normal. Among those in-

ted directly into the same hypotonic solution, 8% of the platelets could be considered deformed, three of those 8% showed adjoining balloons, and 5% had developed fine, long protrusions. The remaining 92% of the platelets were still with spicules, and were counted as normal although they were very swollen.

At ionic strength 0.066 N (40 minutes dialysis) the percentage of the swollen though normal platelets was 92%, the remaining 8% being thick, rounded platelets of smaller diameter having no spicules. Of the platelets that were treated by the drastic method to the same ionic strength, only 58% were swollen normal cells. The rest of the drastically treated platelets had attached balloons, or were thick, rounded platelets of smaller diameter without spicules, some of them with swordlike protrusions.

The greatest difference between the gradual and the drastic method occurs at an ionic strength of 0.058 N (50 minutes dialysis). Of the platelets in the dialysis bag 72% still had spicules, which were slightly longer than those of the platelets in an isotonic solution, but otherwise looked normal, about 20% were thick, rounded platelets and the rest were platelets with attached balloons or with a swordlike protrusion. At the same salt concentration, drastically achieved, only 10% of the cells were morphologically normal, 34% were swollen up like balloons, 10% had developed swordlike protrusions, and 53% were thick, rounded cells.

When the ionic strength reached 0.050 N (60 minutes dialysis) only 25% were morphologically normal although somewhat irregular. Of the rest, 42% were platelets with attached balloons, 3% had swordlike protrusions, and 30% were thick, round platelets.

At the same solution in the drastic method only 5% of the platelets were morphologically normal. Sixty-one per cent were platelets with balloons and 37% thick, rounded platelets.

In the next solution (0.043 N) almost no difference could be observed under the phase-contrast microscope between the platelets in the dialysis bag and those inserted directly into the same hypotonic

solution. In both cases about 87% of the platelets were balloons and 13% of them were rounded thick cells.

This comparative enumeration is summarized in figure 1 where the percentage of morphologically normal platelets in the gradual method is compared to that of the "drastic" method.

Electron microscope observations. These generally confirmed the results obtained by the phase-contrast microscope about the percentage of remaining normal cells at various salt concentrations. At an ionic strength of 0.066 N, however, a major difference, not observed in the phase-contrast microscope, becomes evident between the electronmicrographs of thrombocytes subjected to the gradual and drastic decrease in salt concentration respectively. A great number of torn and otherwise deteriorated membranes (figs. 4, 5) result in the drastic method, while very few of such damaged membranes appear in the gradual method in which the predominant picture is that of swollen but not torn thrombocytes (fig. 3). Another striking feature in the electronmicrographs is the evident increase in the size of the thrombocytes as the ionic strength goes down (fig. 6), and the recovery of the original size when the thrombocytes are brought back into physiological solutions (fig. 7). The damage to thrombocytes that had been brought down in ionic strength drastically (fig. 8) and then returned to physiological saline is much more apparent (fig. 9) than the damage caused to thrombocytes that were treated by gradual decrease in salt concentration (fig. 7). The method used for bringing back the thrombocytes to the isotonic medium, i.e., either dialysing the platelets in hypotonic medium against isotonic NaCl solution or adding sufficient amount of hypertonic solution did not make a notable difference. It should be noted that while the reversed cells from the gradual decrease did not look exactly like the original normal cells in isotonic solution, they recovered their spicules and normal size (fig. 7). The cells that were subjected to a drastic decrease in salt concentration rarely recovered their spicules and mostly were deformed (fig. 9).

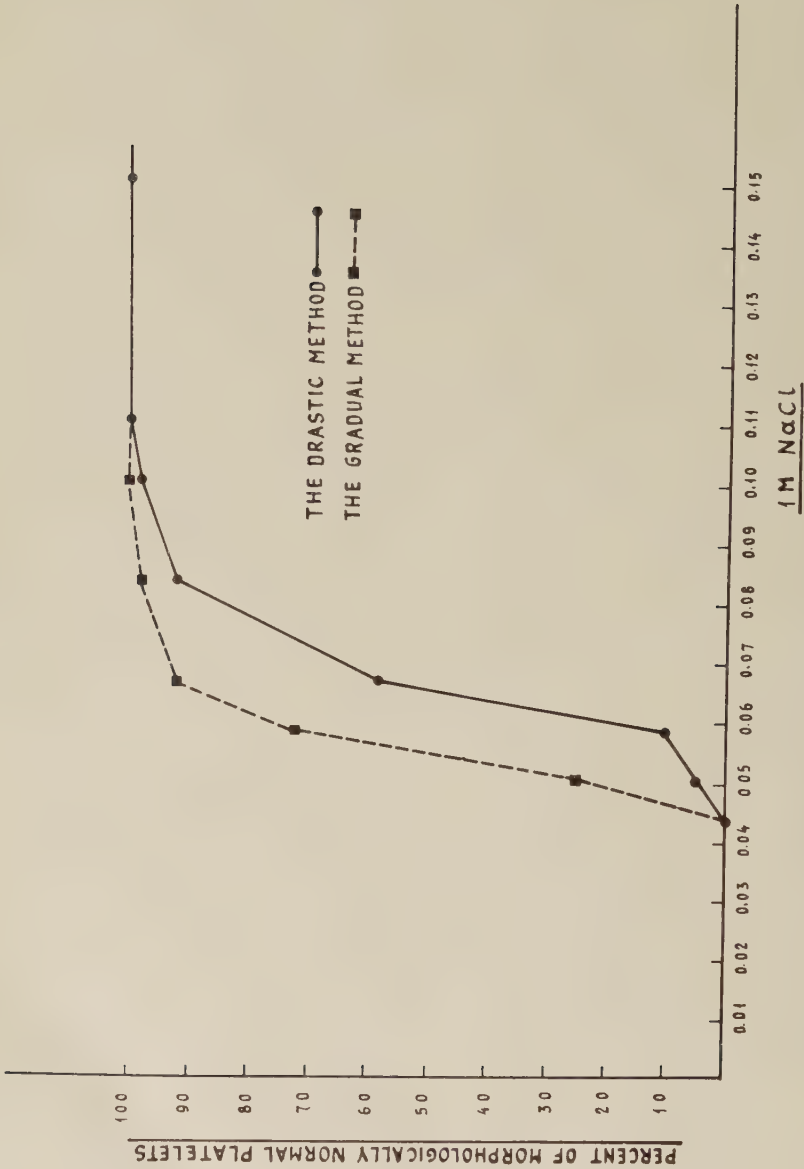


Fig. 1 The number of morphologically normal platelets left after decreasing the salt concentration by the "drastic" and gradual methods.

II. Determination of pyrophosphatase activity

Differences between the gradual and drastic decrease in salt concentration treatments appeared also in pyrophosphatase release from the thrombocytes. While there was no release of pyrophosphatase from the platelets in the dialysis bag at an ionic strength of 0.043 N NaCl, a point at which there was no longer any difference between the two methods in the phase-contrast countings (fig. 1), there was a considerable release of the enzyme from the platelets inserted directly into hypotonic solution of this concentration.

III. Clot retraction experiments

After bringing back the thrombocyte to isotonic solution, their capacity to cause clot retraction was tested. It was found that no clot retraction activity was observed by the thrombocytes treated by the drastic method while the cells treated by the gradual method caused a partial clot retraction. The method used for bringing back the thrombocytes to the isotonic solution, by dialysis or by adding hypertonic solution, did not make any difference in the clot retraction capacity of the thrombocytes.

DISCUSSION

The phase-contrast as well as the electron microscope indicates that there is no basic morphological difference between platelets that have been subjected to sudden decrease in the salt concentration of the surrounding medium, and those that have reached the low salt concentration very gradually by the dialysis method. Apart from details, the modifications of the platelets' shape are similar in both methods. The percentage of thrombocytes of various shapes, however, is different for the two methods at corresponding salt concentrations. A certain fraction of the cells treated by the drastic method, moreover, present a broken membrane. This is demonstrated on the one hand by the electron micrographs showing the damaged and broken membrane and on the other hand by the leakage of pyrophosphatase. Another significant difference between the two methods appears when the ionic strength of the suspending medium is re-

versed to the physiological. Here, we see that the drastic method caused a total loss of the capacity of the platelets to bring about clot retraction, while the platelets that were treated by the gradual method partially conserve this capacity. The results obtained by the gradual method indicate that the membrane can endure large deformations without irreversible changes as long as these take place gradually. The reversibility of the platelet to a shape similar to its original as well as the conservation of its capacity of clot retraction is an indication of the rubber-like nature of its membrane. The molecular mechanisms in the membrane will be loaded by the stress and their ability to withstand the stress applied will depend on the rate of stressing. It seems that a certain time is necessary in order to bring about architectural changes on the molecular level in the membrane without breaking it. If this time is not available, as in the case of rapid swelling, the changes will be accompanied by disruption of the inter-molecular structure. Such behavior has already been attributed to the erythrocyte membrane (Klibansky, '60). In the thrombocyte, however, the return of the deformed membrane occurs over a wider range. The influence of the time during which the pressure is applied is also more apparent in the case of the thrombocyte membrane.

SUMMARY

Normal human thrombocytes were exposed to gradual and "drastic" decrease in the salt concentration of the surrounding medium. Morphological changes of the thrombocytes were observed in the phase-contrast microscope and the numbers of morphologically normal cells remaining in the various hypotonic salt concentrations were counted and plotted as a function of the ionic strength. Electron micrographs are presented to illustrate the morphological changes caused by the two different methods. The release of pyrophosphatase enzyme is compared for the two methods as well as the clot retraction after return to physiological solutions. It was found that morphological changes resulting from the gradual method are reversible but that irreversible changes are produced by the "drastic" method. Pyrophosphatase is re-

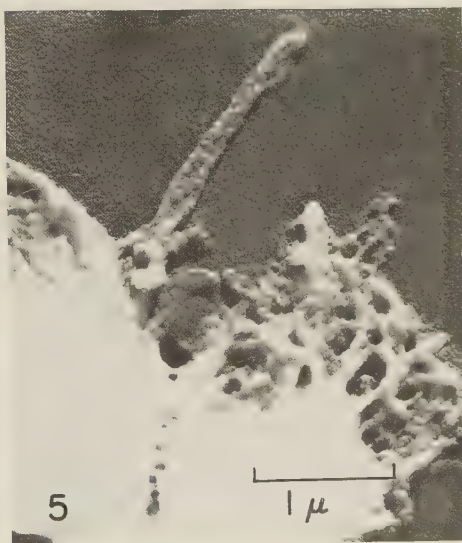
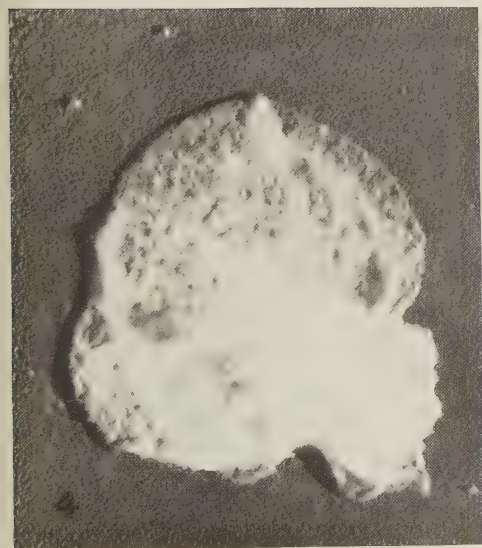
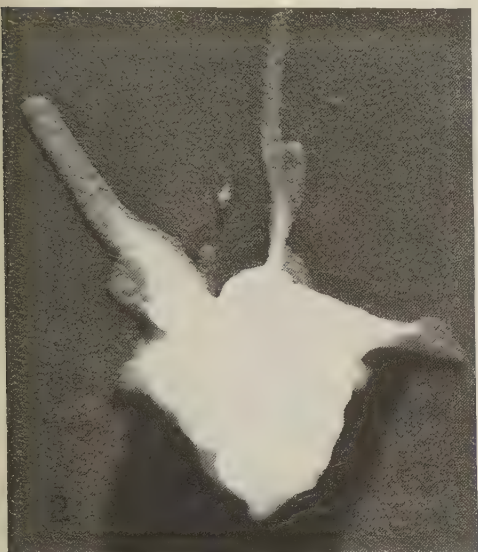
leased only in the "drastic" method and partial clot retraction occurs only after return to the physiological condition in the gradual method.

ACKNOWLEDGMENT

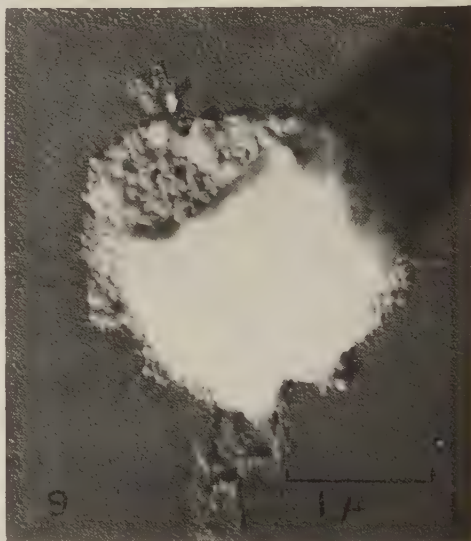
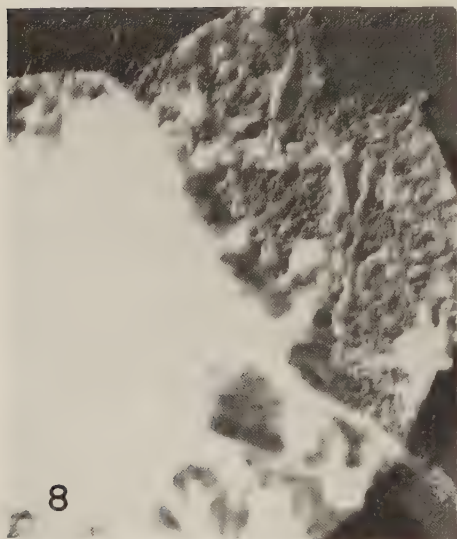
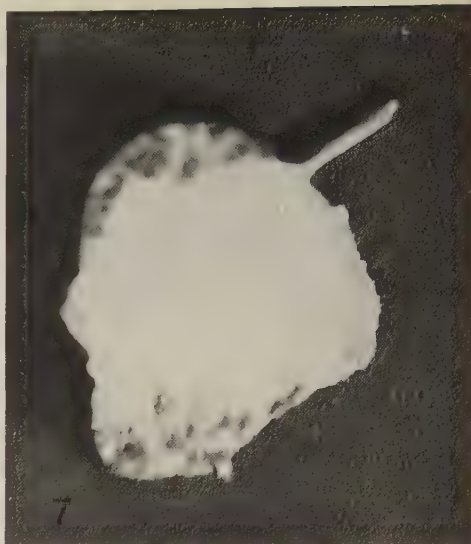
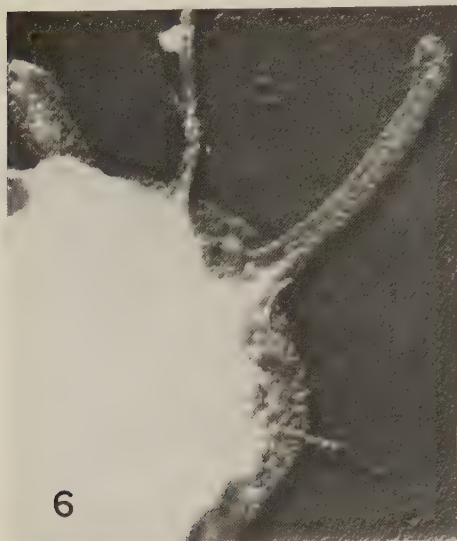
The authors are indebted to Dr. O. Kedem and Dr. A. Silberberg for valuable discussions.

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- 2 Normal human thrombocyte in 0.15 N NaCl.
- 3 Human thrombocyte after gradual decrease in salt concentration of the surrounding medium down to 0.066 N NaCl (50 minutes dialysis).
- 4 Human thrombocyte after sudden ("drastic") transfer into hypotonic solution of NaCl 0.066 N showing an adjoining balloon.
- 5 Same treatment as in figure 4 showing tears and holes in the thrombocyte membrane.



- 6 Human thrombocyte that was gradually brought by dialysis to the ionic strength of 0.043 N NaCl. Note the remarkable swelling.
- 7 Human thrombocyte that was gradually brought, by dialysis, to ionic strength of 0.043 N NaCl and then brought back to 0.15 N NaCl by adding hypertonic solution.
- 8 A thrombocyte brought down to the same ionic strength as in figure 6 but in a "drastic" method. Note tearing of the membrane.
- 9 A thrombocyte that was brought down by the drastic method and then back to physiological concentration. Note the deterioration of the membrane has persisted.

The Electrical Activity of the Amphibian Lymph Heart

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It is well known that the lymph hearts of Amphibia lack the automatism characteristic of vertebrate cardiac tissue. Their activity, like that of Arthropod hearts, depends upon the integrity of their nerve supply. In fact, each contraction of the lymph heart is elicited by a volley of nerve impulses that originate at a small group of spinal motoneurons (see Gada, '56).

The electrical activity of the posterior lymph hearts of the frog was studied by Brücke and Umrath, and also by von Holst, in 1930. Recording directly from the lymph heart muscle these authors observed that each systole is accompanied by a burst of electrical activity consisting of 8 to 10 action currents with an overall duration of 0.2 to 0.5 seconds. The amplitude of the recorded potentials was very low, presumably, to the shunting effect of the connective tissue in which the lymph hearts are embedded.

Brücke and Umrath studied also the spontaneous electrical activity of the XI spinal nerve,² the main source of motor innervation to the lymph heart. Bursts of action currents, just above the limit of resolution of the recording system, were shown to occur in this nerve. Although simultaneous electrical recording from the lymph heart and nerve was performed, it could be concluded that the action currents observed in the XI spinal nerve were responsible for the initiation of the periodic outbursts of electrical activity recorded from the muscle tissue.

Since those papers were published, considerable advances have been made in both instrumentation for electrical recording and our understanding of neuromuscular synaptic mechanisms. It was interesting, therefore, to repeat some of the earlier work with the techniques now available.

METHODS

Most experiments have been performed on the posterior, or coccygeal, lymph hearts of the toad *Bufo marinus*, which are comparatively large and flat, have rather thick walls and are situated just underneath the skin. The lymph hearts in the frog *Rana pipens*, were explored, but they seem to have a more irregular shape and are more deeply located. Tree-frogs *Eleutherodactylus portoricensis* (Schmidt), have also been used.

The toads and tree-frogs were anesthetized with urethane (2.5 gm/kg). This procedure results in more lasting preparations than those obtained by decerebration. Dissection was limited in most instances, to exposing the lymph heart by removing the skin and subcutaneous fasciae.

In the toad, the external surface of the lymph heart is surrounded by tough connective tissue which represents a nearly impenetrable barrier to the intracellular micropipettes. These electrodes could only be used on the inner surface of the organ. To do so the dorsal wall of the lymph heart was cut open. This procedure does not seem to interfere with the motor innervation as hearts prepared in this fashion went on beating vigorously for several hours.

Although the temperature of the laboratory where these experiments were performed was maintained between 19° and 25°C, the observed rates of the lymph hearts, of about 100 systoles per minute, were higher than those reported by other authors for the same temperature range.

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² Called sometimes X spinal nerve, as an embryonic first spinal nerve is missing in the adult animal.

(from 60 to 70 per minute according to Braun-Menendez and Foglia, '40).

Extracellular recording. The electrical activity of the lymph heart was recorded with the aid of a pair of platinum wires (B. & S. gauge no. 24) covered by an insulating coat of varnish. Their exposed tips were applied against the dorsal surface of the heart. No difficulties due to movement artifacts were experienced. The gentle pressure exerted by the electrodes themselves was sufficient to immobilize the heart without producing noticeable damage to the tissues.

The potentials picked up by the platinum electrodes were differentially amplified by a Tektronix type 122 preamplifier and displayed on the screen of a Tektronix type 502 dual beam cathode ray oscilloscope. Photographic records were obtained with a Grass Kymograph camera. The preparation was grounded by means of a large electrode applied to the muscles of the thigh.

Intracellular recording. This was performed with the aid of conventional micropipettes, filled with 3 molar KCl solution and controlled with a Zeiss sliding micro-manipulator. The metal electrode in contact with the KCl solution was connected to the Tektronix 502 CRO via a high input impedance head stage with low grid current and negative capacity neutralization (Lettvin et al., '58).

Recording of motor nerve impulses. Action currents of the XI spinal nerve have been recorded, after removal of the coccygeoiliacus muscle, by suspending the nerve from two platinum hooks in a pool of mineral oil. These electrodes were connected to a Tektronix type 122 preamplifier. As the number of motor axons in the XI spinal nerve supplying the lymph heart is small, not more than about 6 or 7, a high overall gain had to be employed in the recording system. The signal-to-noise ratio could be improved by dissecting from the main nerve trunk a small nerve bundle containing some of the desired motor axons.

Electrical stimulation. The electrical stimulation of the motor nerve fibers supplying the lymph heart could easily be achieved by applying pulses either to the spinal motoneurons or to the XI spinal

nerve. Both these procedures have the advantage of resulting in the stimulation of many other motor axons and the consequent mechanical and electrical artifacts produced by the activity of surrounding muscles. Instead, by exploring the connective tissue surrounding the lymph heart with a monopolar electrode, it was usually possible to find a site where weak electrical stimuli gave rise to a contraction of the heart without involving the adjacent skeletal muscles.

Recording of lymph heart movements. The movements of the intact lymph heart have been recorded by a method developed in collaboration with Dr. J. Y. Lettvin. It takes advantage of the fact that the mechanical distortion of the tip of a high resistance glass micropipette results in marked changes in the electrical impedance between the inside of the pipette and the external solution. So a micropipette was placed upon the heart in such a position that its tip moved and bore with each systole. The impedance of the micropipette tip was continuously monitored by applying rectangular pulses of current of constant strength and duration of 2 msec. at a frequency of 20 cycles/sec. The voltage drop produced by the current flow, which is proportional to the impedance of the tip, was displayed on the oscilloscope using a relatively slow sweep speed. An envelope was thus obtained representing the variations of the micropipette impedance. It is obvious that either an increase or a decrease of surface impedance may accompany each systole depending upon the initial position of the micropipette tip and the pressure applied. In figure 8 the micropipette was arranged so that the contraction of the heart produced an increase in tip impedance represented by an increase in the width of the envelope.

RESULTS

Extracellular recording from the lymph heart muscle

The records obtained with extracellular electrodes confirmed the results reported by Brücke and Umrath ('30). Bursts of electrical activity made up of a variable number of action currents, are seen associated with each lymph heart contraction.

both recording electrodes are placed on the lymph heart these action currents appear as diphasic variations (see fig. 1) whose amplitude varies considerably not only in different preparations, depending on the magnitude of the external shunt, but also from burst to burst in the same heart. The relative size of the positive and negative components of each action current are also variable and depend mainly on the position of the electrodes on the dorsal wall of the lymph heart. The only

nearly constant feature that one finds in these records is a somewhat slower monophasic wave at the end of each discharge, whose polarity is opposite to that of the main components of the diphasic currents.

The patterns formed by these action currents become simpler, more reproducible, and easier to interpret if only one of the recording electrodes is left in direct contact with the lymph heart muscle. The other lead may be placed on either a region of the heart previously damaged by

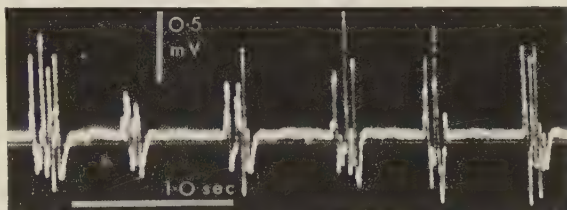


Fig. 1 Bursts of action currents recorded from a lymph heart by means of two platinum electrodes in contact with its dorsal surface. A heart systole is associated with each burst (see fig. 9). Notice the variability in the amplitude of the recorded currents and the duration of the intervals between consecutive systolic discharges.

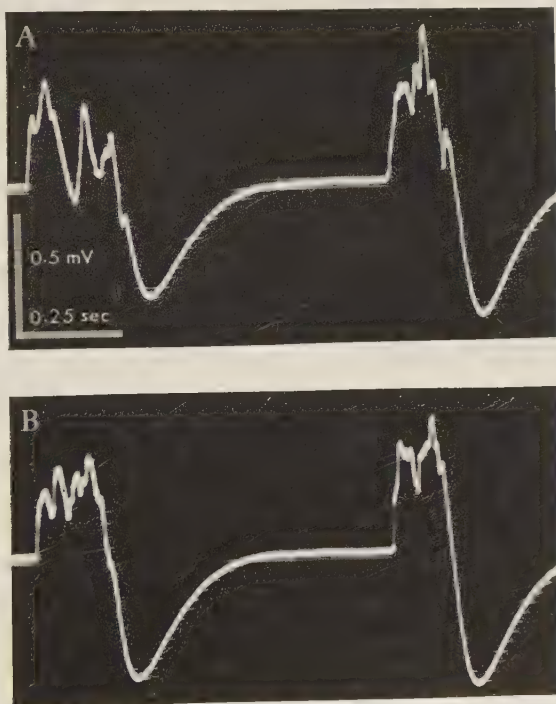


Fig. 2 Systolic discharges recorded from a lymph heart with external electrodes. Whereas in figure 1 *both* electrodes were in contact with the heart muscle, the above records were obtained by placing one electrode directly on the heart and the other on the surrounding connective tissue a few millimeters away. Calibrations in A apply also to B.

local heating, as Brücke and Umrath ('30) did, or in the surrounding connective tissue as we have done in most experiments.

In many preparations, the systolic discharges recorded in this fashion consist of two separate phases (see fig. 2). The first one is made up by a variable number of relatively fast waves (usually 5 to 10) that add up to produce an electrical variation of serrated or spiky contour. This is followed by a slower wave of smooth outline and opposite polarity whose amplitude is comparable to that of the first phase. It must be emphasized, however, that in some lymph hearts we have been unable to obtain such a clear-cut pattern of systolic discharge as those shown in figure 2. In those instances (see fig. 3) the recorded patterns seem to be intermediate between those shown in figures 1 and 2.

The polarity of the systolic discharges recorded with a couple of external electrodes in contact with the dorsal wall of the lymph heart depends upon the relative position of their tips. If the electrodes are radially oriented, the middlemost electrode becomes positive with reference to the

peripheral one during the first wave of each diphasic action current; indicating that this deflection is produced by current flowing from the center to the periphery of the heart.

Similar results have been obtained with a single "active" electrode is placed upon the center of the lymph heart. The relatively fast peaks that form the initial portion of the systolic discharges are due to the transient positivity of that electrode with reference to the "remote" electrode situated outside of the lymph heart area.

If an intracellular microelectrode is inserted into the lymph heart muscle cells (see later) one sees that the initial waves of the systolic discharge represent brief depolarizations of their surface membrane. On the other hand (see last section Results), the externally recorded action currents resemble the first derivative of those potential changes.

This suggests that the externally recorded positive deflections just discussed are due to current leaving the lymph cardiac muscle cells across passive areas of membrane; i.e., regions that do not participate in the permeability changes responsible for cell depolarization.

Although no attempt has been made in this study in detail the distribution of sinks and sources of current in the lymph heart sites have occasionally been found near the edge of its dorsal wall, where action currents of a polarity opposite to that recorded in figures 1 and 2 could be recorded. Exploring those sites with the platinum electrodes one could see that small displacements of the electrode tip may result in a reversal of the sign of the recorded currents (see fig. 4). In between "positive" and "negative" areas the electrode passes over points where the amplitude of the action currents appears greatly reduced.

Electrical activity of the XI spinal nerve

As we have mentioned, Brücke and Umrath ('30) demonstrated the existence of spontaneous bursts of action currents in the XI spinal nerve of the frog which occur at the same frequency as the lymph heart systoles. The lymph heart contractions stop as soon as the nerve supply

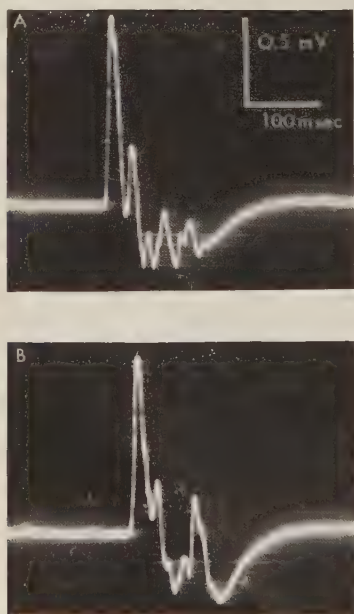


Fig. 3 Two systolic discharges recorded from a lymph heart with the same electrode arrangement as that used in figure 2 (see text). Calibrations in A apply also to B.

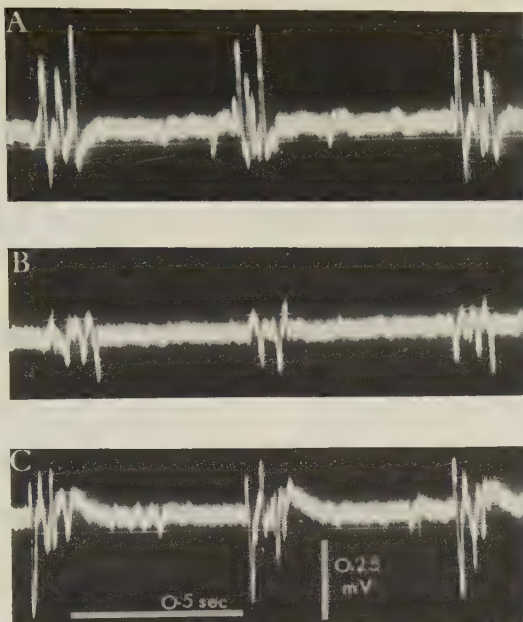


Fig. 4 Systolic discharges recorded from the dorsal surface of the lymph heart with a thin platinum electrode (0.1 mm in diameter). Its position was shifted from record A to B, and from B to C, in two steps of about 0.7 mm each. Notice changes in the patterns formed by the action currents (see text). Calibrations in C apply to all records.

interrupted although discharges of action currents go on in the central portion of the cut nerve. This indicates that the spontaneous nerve activity represents the alleys of motor impulses which elicit each systole. It was, therefore, interesting to record simultaneously the electrical activity of both nerve and heart to observe more closely their relationships.

For this purpose, the electrical discharges of both nerve and lymph heart were picked up by platinum electrodes and displayed on separate beams of the CRO. It could then be seen how each burst of muscle action currents is associated with a train of nerve spikes.

If the entire nerve trunk is suspended from the electrode, the amplitude of the motor spikes is barely higher than the baseline noise. However, it is possible to increase the relative magnitude of the signals by dissecting small bundles of fibers from the XI nerve placing them separately on the recording electrodes. In this fashion preparations can be obtained containing a few lymph-cardiac motor axons which give relatively large spikes. Records

made from these preparations are illustrated in figure 5. The correspondence between nerve and muscle activity is very obvious in some systolic discharges. In record 5F, for instance, each muscle action current is associated with spike activity in the bundle of fibers placed upon the nerve electrodes. In some records such as 5D, a certain correlation seems to exist between the number of nerve spikes and the amplitude of the corresponding muscle action currents. In record 5A, however, we can see a muscle action current (the first one) which is not accompanied by motor nerve impulses. Conversely, nerve spikes are seen which are not followed by muscle action currents. This shows that the nerve impulses observed are only a fraction of those that arrive at the lymph heart, while the recorded muscle action currents represent the activity of only a part of the organ. It is clear, however, that each fast deflection recorded from the muscle is preceded, and presumably elicited, by a variable number of motor impulses reaching the lymph heart along several axons. These impulses are not

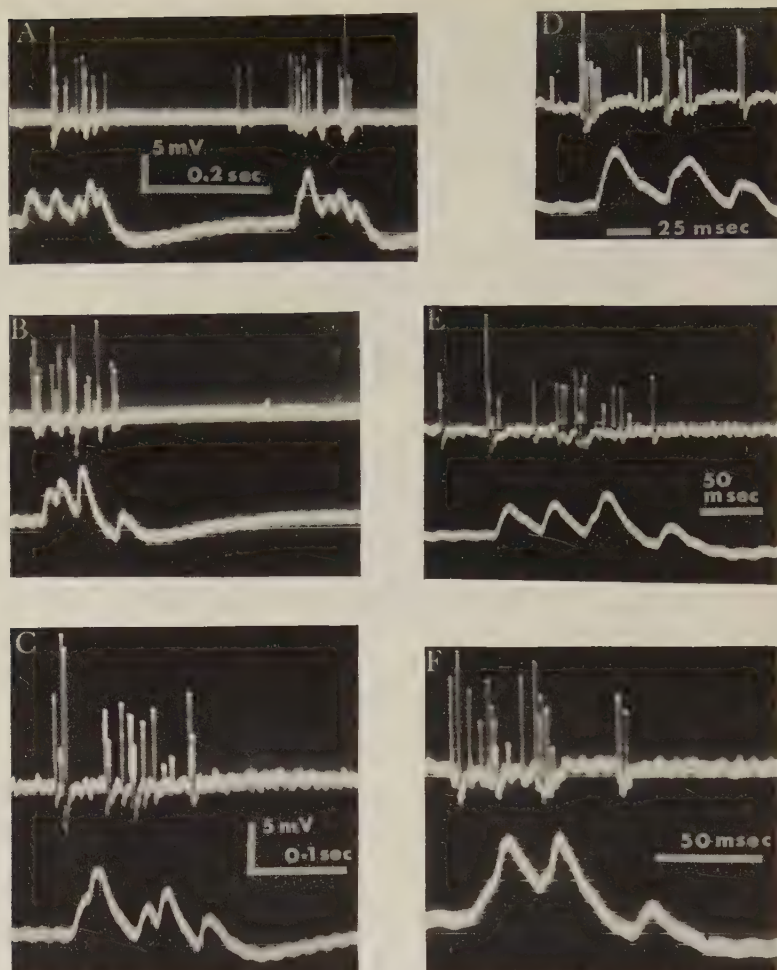


Fig. 5 Simultaneous recording of lymph heart muscle action currents (lower trace) and spontaneous electrical activity in a bundle of nerve fibers dissected from the XI spinal nerve (upper trace). The nerve spikes represent the motor impulses responsible for the synaptic activation of the lymph heart muscle. Voltage calibration in record A applies also to B, D and E, and that in record C applies also to F (lower traces only) time calibration in A applies also to record B.

synchronous, although all of those associated with a single muscle action current seem to arrive at the lymph heart within a period not longer than 12 to 15 msec., see for instance record D in figure 5.

Electrical stimulation of lymph-cardiac motor axons

Another way of obtaining information on the relationship between nerve and muscle activity at the lymph heart is to stimulate directly the lymph-cardiac motor

fibers. The easiest method to do this is to explore the surrounding connective tissue with a monopolar stimulating electrode. Weak negative electric pulses are applied between the tip of this electrode and a larger one making contact with the muscles of the thigh.

Although the anatomical course followed by the motor axons is variable, sites could be located in most preparations where the applied pulses gave rise to contractions of the lymph heart not accom-

nied by twitching of adjacent skeletal muscles.

Figure 6A illustrates the effect of a single supramaximal pulse applied to the lymph-cardiac motor axons. The stimulus, masked by the artifact, is followed after a brief delay by a diphasic action current consisting of a relatively fast initial phase continued by a slower deflection of opposite polarity. These two phases will be referred to as *fast* and *slow*.

The overall duration of the fast phase, from its beginning to the point where the baseline is crossed, is of about 40–45 msec.; the time of rise to peak accounting for approximately one third of this period. The leading edge of the fast wave often exhibits some irregularities which might be due to differences in the excitation + induction time of the various motor axons.

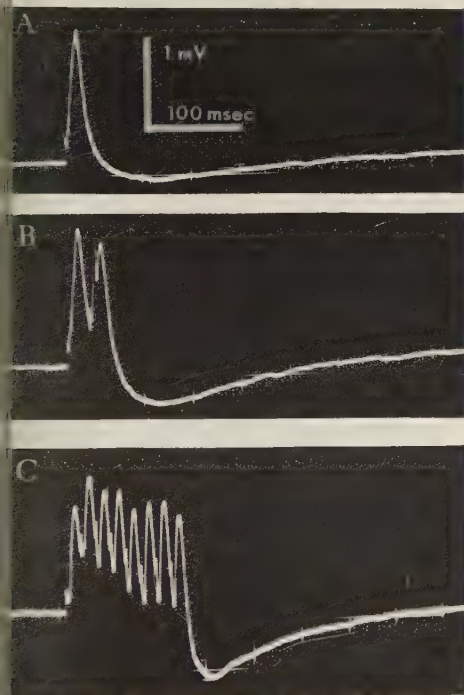


Fig. 6 Action currents of the lymph heart muscle elicited by direct electrical stimulation of the nerve supply (see text). In A a single stimulus is applied whereas B and C show the effects of two and 8 stimuli respectively. In record B the interval between the two stimuli is of 25 msec. The intervals between the stimuli in record C are 20 msec. Calibrations in record A apply also to B and C.

The overall duration of the second or slow phase is of over 0.25 sec. (at a temperature of 23°C); that is, about 6 times longer than the fast or initial phase. The time of "rise" of this phase, to a rather ill-defined peak is of about 75 msec. at the same temperature.

If two pulses are applied in rapid succession (see fig. 6B), the double peak formed by the partial summation of the fast waves elicited by each stimulus is followed by a single slow wave of the opposite sign. No indication of the individual "slow" phases can be distinguished as steps or notches; the single slow wave shows, indeed, a very smooth and uninterrupted contour.

Figure 6C illustrates the effect of applying 8 consecutive stimuli to the lymph heart motor fibers. The resulting pattern of action currents is very similar to some of the spontaneous systolic discharges. Another example of fast repetitive stimulation of lymph-cardiac motor fibers can be found in figure 8B.

Figure 7 has been drawn by superimposing tracings of separate records showing the lymph heart action currents elicited by 1, 2, 3, 4 and 5 stimuli applied to the motor axons. The interval between consecutive stimuli was of 20 msec. The insert shows the maximum amplitude of the second or slow phase in each record plotted against the number of applied stimuli. It can be seen that the amplitude of the slow wave following 1, 2, 3, or 4 fast deflections is equal to the sum of an equivalent number of slow waves following single stimulus.

The polarity of the action currents elicited by direct electrical stimulation of the lymph heart nerve supply depends also upon the position of the "active" recording electrode within the lymph heart area. This polarity is identical with that of the spontaneous systolic discharges recorded with the electrode in the same position. This can be shown by stimulating the motor axons in preparations with an intact central nervous system and lymph-cardiac nerve supply. By doing so, it is possible to record side by side, the muscle action currents generated by motor impulses originating at lymph-cardiac spinal motoneurons and those elicited by the

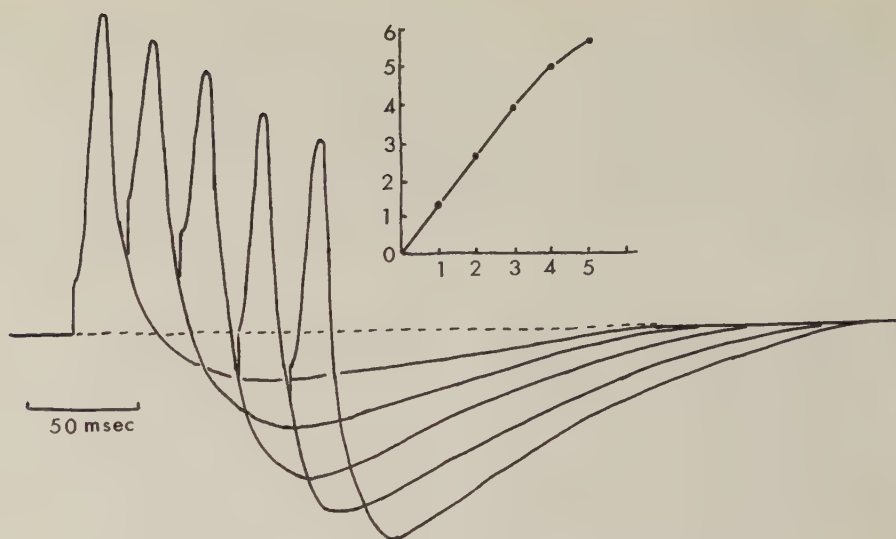


Fig. 7 This figure has been drawn by superimposing tracings of separate records showing the lymph heart action currents elicited by 1, 2, 3, 4 and 5 stimuli applied to the motor axons at a frequency of 50 per second. The insert shows the maximum amplitude of the slow phase in each tracing plotted (ordinate) against the number of applied stimuli (abscissa).

electrical stimulation of their axons in the periphery. This is illustrated in figure 8.

Relationship between the electrical and mechanical activity of the lymph heart

It is easy to see that each systole of the lymph heart is associated with a burst of electrical activity. However, to establish more precisely the relationship between these two events, experiments have been performed in which the movements and the electrical activity of the lymph heart were displayed separately on each beam of the CRO.

By recording the changes of the impedance of a micropipette applied against the dorsal surface of the exposed lymph heart an overall picture of its movements can be obtained. The records resulting from these experiments, some of which are illustrated in figure 9, show that the amplitude of the displacements of the lymph heart tissue is not constant. Very often, furthermore, systoles are observed in which the rising phase of the contraction is interrupted by a step or even by a brief period of relaxation. These patterns are invariably associated with a clustering of the corresponding action currents into two dis-

tinct groups separated by a brief silent period. However, when the action currents form compact patterns such as those seen in figure 9, the ensuing contractions have smooth and uninterrupted contours and single well-defined peaks. The foot, or beginning, of the contraction appears approximately 23 msec. (average of 4 contractions) after the initiation of the systolic discharges. The peak mechanical displacement occurs generally at the end of the systolic discharge, just before the recorded potential returns to the baseline.

After the peak of the contraction is reached, relaxation takes place at a rate that soon reaches maximal value and decays before reaching the presystolic level. In most records the relaxation proceeds until the beginning of the following systolic contraction.

Action of some drugs upon the lymph heart

The influence of different compounds upon the lymph heart has been studied by several investigators; their findings, summarized by Braun-Menendez and Fogli ('40) (see also Priestley, 1878, for references to early work), show that this organ is particularly sensitive to drugs acting on

linergic junctions, such as nicotine, mscarine, d-tubocurarine (dtC), acetylcholine (ACh) and other related substances.

We have repeated some of those experiments and have confirmed the main results. Our observations have shown, however, that drugs are active on the lymph heart at concentrations significantly lower than those reported in previous papers. For instance, dtC said to block the lymph heart contractions at a concentration of 10^{-2} w/v (see Foglia and Braun-Menendez, '39) is still active if applied locally at concentrations of 10^{-5} to 10^{-4} w/v. ACh is also active at similar concentrations. This discrepancy can probably be explained by

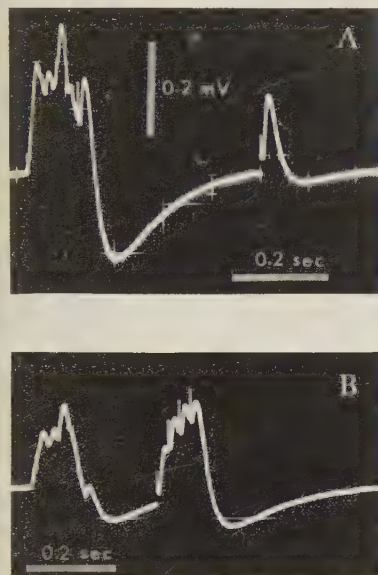


Fig. 8 Records showing systolic discharges generated by spontaneous neural activation of the lymph heart together with action currents elicited by electrical stimulation of lymph-cardiac motor units in the periphery. In A, an action current induced by a single shock applied to the motor units (easily recognizable by the stimulus artifact) appears, by chance, at the end of a spontaneous systolic discharge made up by at least 5 distinct peaks. A single action current elicited by electrical stimulation is seen in the same sweep. Notice the marked difference in the amplitude and time course of the two action currents elicited by artificial stimulation (see discussion). In B, a spontaneous systolic discharge made up by at least 5 action currents occurs just before a rather similar pattern elicited by 4 consecutive electrical stimuli. Voltage calibration in record A applies also to B.

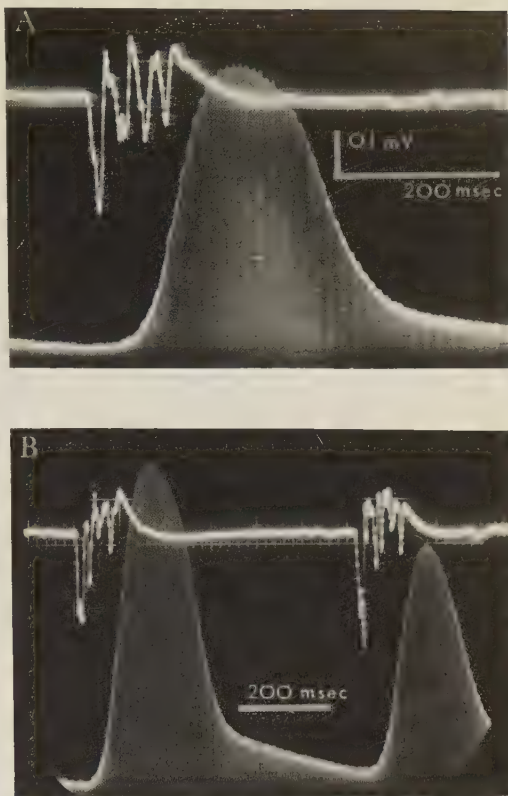


Fig. 9 Simultaneous recording of the electrical activity (upper trace) and movement (lower trace) of the lymph heart muscle during spontaneous systolic contractions. Voltage calibration in A applies also to B.

the fact that we have applied the drug solutions directly on lymph hearts after cutting open their dorsal wall. If the lymph heart is intact, the diffusion of the compounds into the muscle tissue is likely to be hindered by the very dense connective layers which cover its dorsal surface. This difference in technique might also explain that although atropine at concentrations as high as 10^{-2} w/v had been reported to be inactive (Foglia and Braun-Menendez, '39; Hotovy, '39) we have found that a block of the heart contractions is produced by atropine sulfate at a concentration of 10^{-3} w/v.

While working with atropine solutions we have confirmed one of the observations reported by Brücke and Umrath ('30); namely, that a lymph heart blocked by ACh at a concentration of 10^{-4} w/v

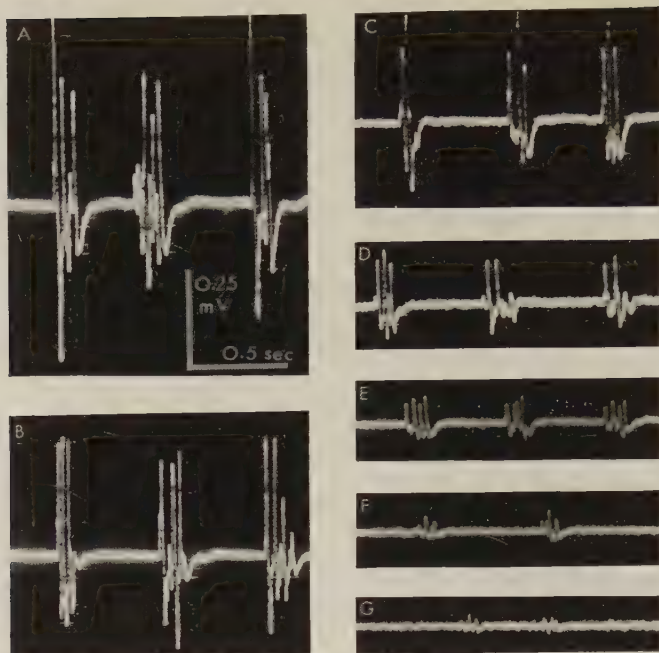


Fig. 10 Effect of d-tubocurarine upon the electrical activity of lymph heart muscle. A, normal systolic discharges used as control. After record A was taken 10 mg/kg of d-tubocurarine were injected intraperitoneally in the toad. Records B, C, D, E, F and G were taken 10, 30, 50, 70 and 80 minutes, respectively, after the injection of the drug. The amplitude of the recorded action currents shows a gradual decrease until they merge into the base line noise. No significant changes in the patterns formed by them can be distinguished (see text). Time and voltage calibration in A applies to all records.

starts beating again if atropine, at the same concentration, is added even if the initial concentration of ACh is maintained throughout the experiment.

The electrical activity of the lymph heart muscle under the influence of dtC and ACh was also examined. Particular attention was paid to the effect of d-tubocurarine on the lymph heart action currents. If the diphasic action current (fig. 6A), that may be regarded as the basic pattern of electrical activity of the lymph heart muscle, represented a combination of an excitatory post-synaptic potential plus a superimposed action potential, one would expect that the block produced by dtC would help to distinguish between those two types of phenomena. These expectations were not fulfilled. If one examines the electrical activity of the lymph heart while a dtC-block is established one sees how the amplitude of the muscle action currents decreases gradually until

they merge into the baseline noise (fig. 10). We have not been able to find either discontinuities or marked changes in the shape of the recorded action currents, such as those observed during curarization of skeletal nerve-muscle preparations in which muscle action potentials give place to end-plate potentials.

Similar results have been observed after the intraperitoneal injection of large amounts of ACh (1 ml of a 2×10^{-2} w/v solution). In other experiments, solutions containing ACh at lower concentrations (10^{-4} w/v) have been applied directly to the open lymph heart while its electrical activity was being recorded. Under these conditions ACh caused not only a marked decrease in the amplitude of the action currents; in some instances the polarity of the systolic discharges was reversed.

These results can be explained on the basis of a differential action of ACh upon the lymph heart cells, due presumably

the existence of diffusion barriers. The more accessible synapses will be blocked first. This will alter the distribution of "sources" and "sinks" of current in the tissue.

It must be emphasized that although a 10^{-4} w/v solution of ACh did not completely block the systolic discharges of the lymph heart currents the corresponding poten-

tial changes in the cell membrane were obviously too weak to elicit an appreciable mechanical response.

Intracellular recording

The lymph heart offers an unfortunate combination of factors that hinder the use of microelectrodes: small cell size, tough connective tissue, and extremely

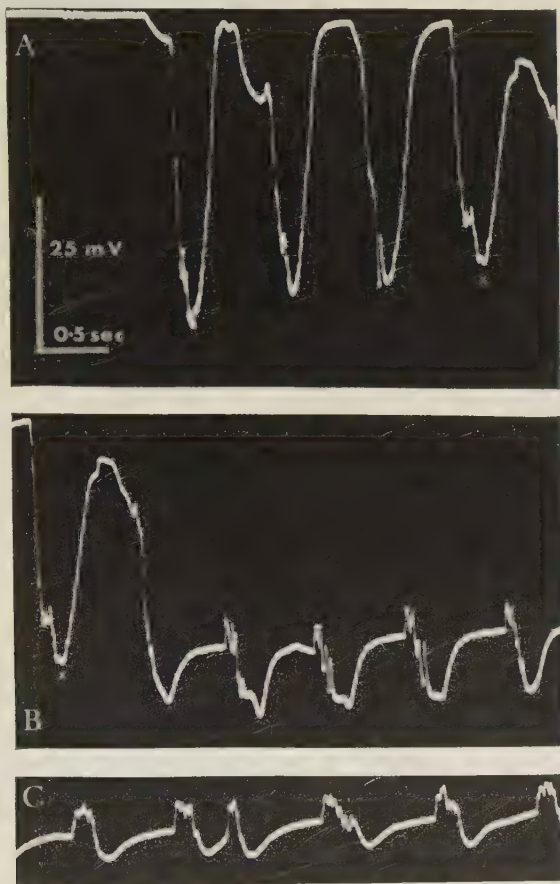


Fig. 11 This figure illustrates the potentials observed during a slow impalement of a lymph heart muscle cell with a micropipette and also the electrical activity of the cell membrane associated with each systole. The microelectrode is shifted steadily towards the muscle tissue from the beginning of record A until the end of B. Both records overlap and the deflection marked with an X in A and B is the same one.

The trace in the upper left corner of A shows the zero level; i.e., the potential recorded when the micropipette tip is in the external solution just outside the cell membrane. The large downward deflections that follow (indicating a negativity of the micropipette) represent transient records of the resting potential due to momentary penetrations of the microelectrode into the cytoplasm of the muscle cell. Eventually, in record B, the microtip stays in the cell and the changes in membrane potential associated with each systole can be observed. The last deflection in record B and those shown in record C are typical systolic discharges as recorded with intracellular microelectrodes (see also figs. 12 and 13). Soon after the end of record B the microelectrode came out of the cell. Record C was obtained during a new impalement in the same site. Calibrations in A apply also to B and C.

vigorous mechanical activity. A number of measures and devices were tried, with only partial success, to counteract these factors. For instance, several species of anura and specimens of different ages have been employed. Proteolytic enzymes (trypsin and papaine) were used in some experiments in order to digest partially the connective tissue although no obvious difference could be observed. Our best results were obtained by inserting the microelectrodes into the inner surface of the heart after limiting its movements by means of a small glass ring (about 1 mm I.D.) applied with gentle pressure. In spite of this, we have obtained only a rather incomplete picture of the electrical activity of the lymph heart membrane.

If the movement of the heart is not completely restricted and the micropipette tip is advanced slowly towards the inner surface of the heart one can see, when contact with the tissue is first established, that the microelectrode tip goes briefly into the cells on each systole. This is shown by the appearance of short-lasting resting potentials coinciding with the heart contractions (see fig. 11A). The magnitude of these brief potentials goes usually up to 45 or 50 mv, although figures of 60 mv have been observed a few times. Their exact value depends, presumably, upon the extent of the cell damage produced by the erratic insertions. We regard, therefore, the largest value (about 60 mv) as a more faithful representative of the real potential difference across the cell membrane than the average value of the smaller resting potentials usually encountered.

If the microelectrode tip is pushed further into the cell, in an attempt to achieve a more lasting insertion, the resting potential falls usually to a value of between 20 and 30 mv. It stays at this low level during a few systoles before disappearing altogether.

More durable insertions are obtained when the pressure exerted by the glass ring is increased until the movements of the tissue within the ring are fully restrained. Resting potentials of between 40 and 50 mv lasting several systoles are often obtained and, in a few instances, values of above 50 mv could be held for a few seconds. It can be seen, in these rec-

ords, that each lymph heart systole is accompanied by a burst of potential changes consisting of several relatively fast components, or "peaks," which represent membrane depolarizations, followed by a slow wave of hyperpolarization (see figs. 11 and 12).

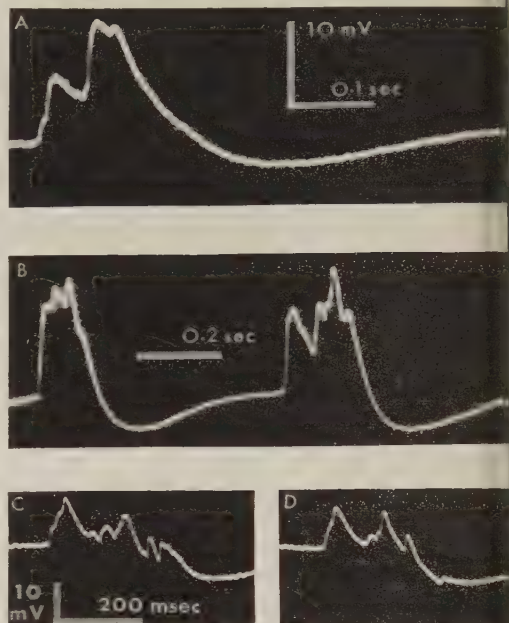


Fig. 12 A, B and C, changes in the potential across the surface membrane of lymph heart muscle cells associated with systolic contractions. The resting potentials on which these potential changes were superimposed measured just over 40 mv. The voltage calibration in record A applies also to B. Calibrations in C apply also to D.

The magnitude of the systolic potential changes depends mainly upon the value of the resting potential on which they are superimposed. When this is low (20 to 30 mv) the amplitude of the "peaks" is only 3 to 4 mv. With resting potentials of 40 mv they may reach up to 15 mv. The largest systolic depolarizations recorded by us had an amplitude of just over 30 mv on a resting potential of 53 mv. In this instance, single peaks of up to 20 mv in amplitude could be distinguished.

In spite of the existence of the above relationship, no attempt has been made to correlate exactly the amplitude of the peaks with the resting potential, as their size also depends upon other factors not

understood. Very often the amplitude of the peaks, rather large immediately after the insertion, was seen to decay very rapidly, well before any appreciable change in the value of the resting potential was observed. In some instances, it was possible to increase the magnitude of the recorded resting potential by pushing the microelectrode a little further into the tissue. This maneuver, however, failed to increase the size of the systolic discharge. The disappearance seems to be associated, therefore, with factors other than the establishment of a shunt around the microelectrode tip, perhaps with the damage produced to the motor nerve endings. It must be emphasized, however, that neither the increase in resting potential when the tip of the microelectrode is pushed further into the tissue, nor the evanescence of the systolic potentials was the result of an increase in the microelectrode impedance and associated changes in the frequency response characteristics of the recording system.

Each single component of the systolic discharge, as recorded with intracellular electrodes, consists of a relatively fast rising phase followed by a slower decay. The time of rise to peak of these potentials varies considerably, from 8 to 23 msec., even within a single burst, as can be seen

in figure 13. We don't know the reason for those differences although they are probably an expression of the different degree of electrotonic distortion suffered by potentials originating at various distances from the recording micropipette.

Each of the fast depolarizations is followed by a slow decay. Occasionally, the interval between two consecutive fast deflections is relatively long, of up to 60 or more msec. In these instances one can see how the slow repolarization of the membrane is continued smoothly by a phase of hyperpolarization, or increased negativity of the cell interior (see records C and D of fig. 12). One is justified, therefore, in regarding the diphasic change in the membrane potential (i.e., a fast depolarization followed by a more prolonged hyperpolarization) as the basic component of the systolic discharge.

As these components occur usually in rapid succession, what we see in the records is the result of their summation. The fast peaks add up one upon another creating a wave of depolarization of variable amplitude and irregular and spiky contour. This wave is followed by a smooth and continuous wave of hyperpolarization in which the contributions of each single component cannot be distinguished. The behavior of the diphasic changes in mem-

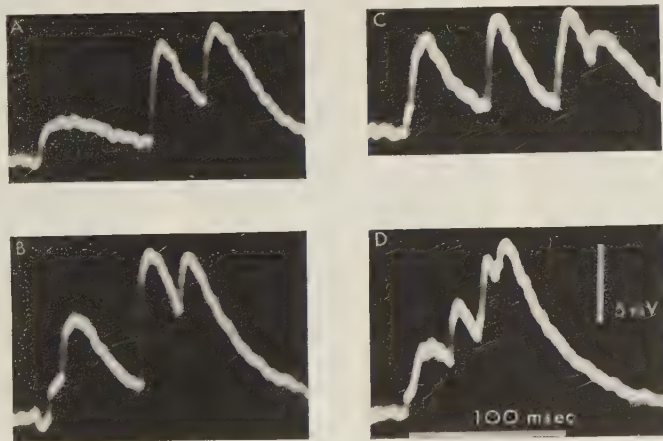


Fig. 13 This figure illustrates the initial phases of 4 systolic discharges of a lymph heart, recorded with a micropipette inserted into a muscle cell. These records were selected because in this cell the initial phase of the discharges was made up of only three or 4 fast components, instead of 5 to 10 as usually found. Notice the large differences in the time course of individual fast waves (see text). Calibrations in record D apply also to A, B and C. The resting potential on which these potential changes were superimposed was of only 35 mv.

brane potential seems to be identical, therefore, with that of the diphasic action currents recorded with external electrodes.

An attempt was made to obtain information on the distribution of the amplitude of the systolic discharge, as recorded with intracellular microelectrodes, over the inner surface of the lymph heart. This proved to be impossible in practice. Although in some preparations sites could be located where potentials of relatively large amplitude were consistently obtained, it could not be ascertained whether the potentials were, in fact, larger there or the conditions for recording more favorable.

Relationship between the systolic discharges recorded with external electrodes and intracellular micropipettes

There is a marked resemblance between the systolic discharges recorded with a single "active" external electrode placed upon the lymph heart and the intracellular potentials just described. In spite of such an obvious similarity (compare, for instance, figs. 2 and 12B) one would like

to find out the exact correspondence between the two events by recording simultaneously with external electrodes and intracellular microtips, from the same site of the lymph heart tissue.

It has been impossible so far, to observe intracellular potentials of the lymph heart muscle cells while the external action currents were recorded with the electrode arrangement used in figure 2. However, in a few exploratory experiments we have been able to pick up intracellular potentials at the same time that the action currents in the vicinity of the microtip were recorded with a couple of thin platinum electrodes separated by a distance of about 1 mm. These electrodes were connected to a differential amplifier whose output was displayed in one of the beams of the CRO. The intracellular potentials were shown in the other beam. Figure 14 illustrates some of the records obtained with this technique. They show that the positive peaks of the *extracellular* action currents coincide with the rising phase of the fast *intracellular* potentials (i.e., depolarizations) while the external negative deflections

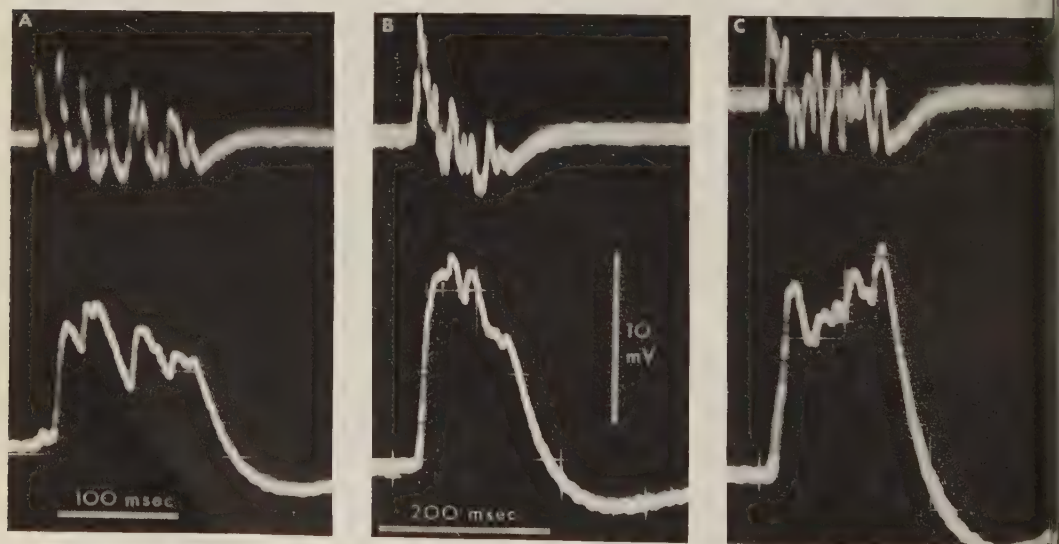


Fig. 14 This figure shows three systolic discharges of a lymph heart recorded simultaneously with a couple of extracellular platinum electrodes (upper trace) and an intracellular micropipette (lower trace). (See text.) The voltage calibration which appears in record B applies also to A and C (intracellular records only). Time calibration in record B applies also to record C. A comparison of the extra and intracellular deflections in this and other records shows that the externally recorded action currents behave approximately like the first derivative of the simultaneous changes in membrane potential.

tions are simultaneous with the re-polarization of the membrane.

Although better records are needed to establish accurately the correlation between changes in membrane potential and ion currents, it seems from the present results that the extracellular action currents behave approximately like the first derivative of the intracellular potentials. This finding is consistent with the assumption that, in most instances, the electrical activity of the lymph heart recorded with external electrodes is due to current flow across areas of membrane that remain passive during the generation of systolic charges. The fast positive deflections observed with external electrodes during the initial phase of the systolic discharge could be produced by current flowing outwards across the cell surface membrane, during the rising phase of the depolarization.

It also appears that the hyperpolarization of the membrane, and the subsequent return of the membrane potential to the initial level, occur at a rate too slow to be detected in an external action current large enough to be detected with the experimental arrangement used to obtain the records of figure 14. However, a careful examination of these records shows that the final and slower, negative deflection that marks the end of the extracellular systolic discharge does not end as soon as the membrane potential reaches the diastolic level, but goes on slightly beyond that point. This short tail may be regarded as a very attenuated extracellular counterpart of the phase of membrane hyperpolarization.

DISCUSSION

The experiments described above show that the systolic discharges of the lymph heart are produced by the addition of a number of diphasic variations of the resting potential existing across the surface membrane of the lymph-cardiac muscle cells. Each of these potential changes, excited by volleys of motor nerve impulses, consists of a relatively fast depolarization, up to 20 mv in amplitude, followed by a slower phase of hyperpolarization. The first question raised by the demonstration of the diphasic lymph heart po-

tentials is that of their nature. In principle, each potential could represent a post-synaptic potential, an action potential, or a combination of both. Their time course does not provide much information on this problem. Although the duration of each of the components of the systolic discharge is long compared with the action potentials recorded from nerve and skeletal muscle of Amphibia, it is short compared with the action potentials recorded from the frog's ventricle (up to 0.8 sec. at 21°C, according to Woodbury et al., '51) or the frog's sinus (with an overall duration of up to 1.6 seconds, at 16°C, del Castillo and Katz, '55). An action potential, however, represents an all-or-nothing event in which the potential difference across the excitable membrane reaches a fixed level. For this reason, action potentials cannot be added in single cells. The intracellular lymph heart potentials, however, add one upon another, suggesting that we are dealing with post-synaptic potentials rather than with action potentials.

Another indication of the synaptic nature of those potentials is the effect of d-tubocurarine on the lymph heart action currents. As figure 10 shows, the establishment of a curarine block is a gradual process not accompanied by noticeable changes in the patterns formed by the action currents.

The experiments in which the influence of dtC, ACh and atropine were studied strongly suggest that the neural activation of the lymph heart depends upon a synaptic mechanism involving the liberation of ACh and its combination with postsynaptic receptors. It is still doubtful whether an enzymatic ACh-inactivating mechanism operates in the neuromuscular junctions of the lymph heart. The results of experiments in which prostigmine at concentrations of 2 to 4×10^{-6} w/v were applied to the lymph heart have so far been negative. It is likely, however, that more refined techniques may be required to demonstrate the effects of anticholinesterase agents.

Potentials resembling the miniature end-plate potentials recorded at the skeletal neuromuscular junctions have occasionally been observed. However, the

uncertainties associated with the measurement of the actual amplitude of the synaptic potentials, emphasized earlier made it difficult to decide whether those potentials were due to spontaneous transmitter liberation or to uncoordinated firing of single spinal motoneurons. Erratic discharges of the spinal lymph heart centers can be observed while asphyxia of the central nervous system develops. When this happens the activity of the lymph heart becomes disorganized before ceasing altogether. The vigorous systolic contractions of the heart are replaced by a weak "fibrillation." Intracellular recording shows then the occurrence of single diphasic potentials not grouped into systolic discharges.

In experiments such as the one illustrated in figure 8A, it can be observed that the time course of single action currents elicited during the phase of hyperpolarization following a previous current is different from those elicited in a resting membrane. Experiments are in progress to determine whether this difference is the consequence of a mechanical artifact or due to a change in the properties of the membrane of the lymph-cardiac muscle cells during the hyperpolarization.

As no action potentials have been observed, it appears that the activation of the contractile machinery of the lymph-cardiac cells is elicited directly by synaptic activity.

Two different aspects should be considered separately when summarizing the above results. In the first place, some of the functional properties of the lymph heart depend upon the patterns of discharge of the spinal centers that supply motor innervation to this organ. Our results in this regard have confirmed earlier work. The records obtained from bundles of fibers of the XI spinal nerve have shown that each lymph heart systole is elicited by a train of motor impulses made up by several (5 to 10) separate volleys. It is likely that each motor axon may contribute one or more impulses to each volley, although the number of action potentials and the duration of each volley, as well as the sequence of fiber discharge, are highly irregular and seem to vary at random from systole to systole.

In the second place, we should consider the intrinsic properties of lymph heart muscle cells. These are characterized, from an electrophysiological viewpoint, by (a) a resting potential which amounts roughly to $\frac{2}{3}$ of that of skeletal muscle fibers, (i.e. about 60 mv, inside negative), (b) the absence of action potentials, and (3) the occurrence of phasic synaptic potentials, in which the depolarization of the cell membrane (up to 20 mv) elicited by a cholinergic mechanism is followed by a longer phase of membrane hyperpolarization. The three properties, together with a diffuse and relatively uniform distribution of nerve endings, are the distinctive features of the slow muscle system of the Amphibia studied by Kuffler and Vaughan Williams ('53; see also Kuffler, '53). Although more work is needed to reach a definite conclusion we may tentatively regard the lymph heart as an organ made of "slow" muscle fibers. We have already mentioned the difficulties encountered when trying to study with electrical methods the pattern of distribution of motor nerve terminals in the lymph heart. It is likely that information on this problem could be obtained more easily from histological studies.

The systolic discharges recorded from the spontaneously beating lymph heart are the result of the summation of a number of diphasic synaptic potentials, each elicited by a separate volley of nerve impulses. Due to the great variability of those volleys with regard to the numbers of impulses, overall duration and sequence of firing of spinal motoneurons the shape, duration and distribution of the systolic action currents are highly changeable affairs.

SUMMARY

1. The electrical activity of the posterior lymph hearts of the toad has been investigated using both external electrodes and intracellular micropipettes.

2. In agreement with previous work bursts of muscle action currents (systolic discharges) were found to be associated with each lymph heart systole. Mechanical activity starts about 20–25 msec. after the beginning of each burst, reaching maximum at the end of it.

If one external electrode is in contact with the dorsal wall of the lymph heart and the other is placed upon the surrounding connective tissue, the first of the systolic discharge appears as a large serrated deflection made of the sum of several (5 to 10) relatively fast repetitive waves. This is followed by a smooth and uninterrupted negative deflection.

If the lymph cardiac motor axons are stimulated with a single electric pulse, a basic action current is recorded from the muscle tissue. Repetitive stimulation shows that the systolic discharge described (3) is the sum of a number of these basic currents.

The electrical activity of lymph cardiac motor axons has been recorded simultaneously with the systolic discharges. The muscle action current is seen to be preceded by a volley of motor impulses. The impalement of lymph heart muscle cells with an intracellular micropipette reveals the existence of a resting potential up to 60 mv in amplitude (cell interior negative).

The systolic discharges recorded with an intracellular microelectrode begin with a number of relatively fast deflections that form a spiky wave of depolarization up to 30 mv in amplitude. This phase is followed by a wave of membrane hyperpolarization of smooth and continuous character.

The intracellular systolic discharge is the same as its extracellular counterpart, the result of the addition of a number of biphasic deflections, i.e., fast depolarizations followed by more prolonged hyperpolarizations. These potential changes may be regarded as postsynaptic potentials produced by a cholinergic mechanism. No action potentials have been recorded from the lymph heart.

The electrical activity of the lymph heart has been recorded simultaneously with an intracellular pipette and a couple of extracellular electrodes placed in the vicinity of the microtip. The external action currents recorded with this electrode arrangement behave approximately as the first derivative of the changes in membrane potentials.

10. It appears from the above results that the electrical activity of the lymph heart muscle resembles very closely that of the slow muscle system of the Amphibia.

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The Possible Role of DNA-ase I in DNA Replication

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It is a well-established fact that native two-stranded DNA which has been isolated from calf thymus cannot itself act as a primer unless it has been denatured (usually by heat) (Bollum, '59), in the synthesis *in vitro* of DNA from the 4 deoxyribonucleoside triphosphates using the polymerizing enzyme from thymus tissue. This two-strand DNA does function as a primer however in Kornberg's system using an enzyme prepared from *E. coli* (Lehman et al., '58). In the latter case a slight degradation of the DNA with DNA-ase increases the rate of reaction two or three-fold (Lehman, '59).

When using an enzyme obtained from perphase liver cells, it has been found that unheated DNA does not act as a primer (Mantsavinos and Canellakis, '59a), but when the enzyme is obtained from regenerating liver, two-strand DNA will act as a primer. In the latter case the addition of DNA-ase I decreases the amount of synthesis at all concentrations investigated. When polymerizing enzyme from certain tumor cells is used, an optimal concentration of DNA-ase I enhances the net synthesis 5-fold (Mantsavinos and Canellakis, '59b). It is thus obvious that the rate of synthesis of DNA from the deoxyribonucleotide triphosphates can be influenced either positively or negatively by DNA-ase action.

Consideration of the results just outlined leads to the suggestion that native two-stranded DNA might need to be converted to single strand material prior to DNA synthesis. This idea is compatible with the fact that the single-stranded DNA which has been isolated from Φ X174 phage by Sinsheimer ('59) can act as a primer for the polymerizing enzyme from thymus (Bollum, '59) or from *E. coli* (Lehman et al., '58). The idea is also compatible with recent work of Zamenhof ('59)

indicating an instability of one of the strands of the two-strand DNA of *E. coli*. However, the question as to whether complete separation of the two strands ("destranding") is necessary for primer action has not yet been settled.

It appears somewhat difficult to imagine a process that could produce material *in vivo* similar to the heated DNA. The idea of completely unwinding the two strands of a double helix prior to DNA synthesis under physiological conditions would seem to present rather formidable difficulties (Delbruck and Stent, '57), although such unwinding or "destranding" may well occur on heating DNA. Hence it seems worthwhile to consider the possibility that single-stranded DNA might be produced as the result of a specific destruction of one strand of the two-stranded helix. It should be noted that either strand of a two-stranded Watson-Crick helix carries all the information needed for the synthesis of more two-stranded DNA.

If destruction of one strand of a DNA double helix were to precede DNA synthesis, the first step in the synthetic process would presumably have to be the exact duplication of this single strand material. Then the single strands could conceivably bind the complementary bases to themselves according to the A-T, G-C rule, and finally after polymerization of the newly bound bases, two double strands of DNA would have been formed for each double strand originally present. This second step in the synthesis could terminate the process according to the suggestion of Bollum ('59), if some way could be found to block further conversion of double to single strand material. A mechanism of this sort might or might not be compatible with the

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results of Meselson et al. ('57) on isotope distribution in DNA after replication.

Since DNA-ase I is capable of enhancing the primer activity of two-strand DNA in certain cases, as stated above, it seemed logical to determine whether DNA-ase I action on the nucleoprotein complex of cell nuclei might lead to the production of single-strand DNA. The reasoning behind this proposition is that one of the fibers of the two-stranded DNA in the nucleoprotein might be sterically hindered by protein, especially by histone, and thus be inaccessible to DNA-ase action.

It was considered of interest to investigate this possibility by studying the action of DNA-ase on cell nuclei isolated at pH 6.0 using very dilute citric acid as a medium. In this case DNA-ase I is liberated from cytoplasmic granules and enters the nuclei during the homogenization and afterwards, causing degradation of the DNA (Dounce et al., '57). We have found that a mild but definite degradation of the DNA occurs if the pH is kept between 5.8 and 6.0, whereas a more extensive degradation takes place if the pH is allowed to rise to 6.1 or 6.15. Above pH 6.2 the nuclei are disintegrated. The two types of DNA in question will subsequently be referred to as *slightly degraded DNA* and *considerably degraded DNA*, respectively. The viscosity of the *slightly degraded DNA* has already been reported (Dounce et al., '57), while that of the *considerably degraded DNA* was about 1/5th that of calf thymus DNA in 0.01 M NaCl solution.

The immediate object of our investigation was to isolate DNA from the already isolated liver cell nuclei and then determine whether the degradation was entirely the result of simple depolymerization (i.e., transection of double helices) or whether a tendency towards production of single stranded material might be noted. We have elected to use the action of DNA-ase I in the homogenate on the whole nuclei rather than to study the action of crystallized DNA-ase I on isolated nucleohistone in order to keep conditions as close to physiological as possible. Isolated DNA nucleohistone may represent a more or less disrupted structure, the study of which might in this case be more apt to lead to erroneous conclusions than the use of the more

complicated system involving the whole nuclei where the DNA-ase action on the DNA takes place as it were *in situ*. We know that histones remain in the pH 6.0 nuclei (Monty and Dounce, '57) and we know that the chief enzyme degrading DNA is DNA-ase I (Dounce et al., '57; see also their references).

In order to test for the production of single strand material, the formaldehyde reaction of Fraenkel-Conrat ('54) was employed, using spectral shift as a measure of the reaction. In addition, studies of base ratios of the isolated DNA were made by the method of Wyatt ('55).

Studies of base ratios of course will not show whether there has been separation of the two strands of the DNA double helix, provided that both strands remain in the sample. If, however, there has been partial destruction and loss of one of the two strands, analysis for base ratios would be expected to show some variance from the A-T, G-C rule.

For purposes of comparison, some studies were also made of calf thymus DNA. DNA isolated from whole rat liver, and DNA prepared from rat liver cell nuclei previously isolated at pH 4.0 (where the DNA is not subject to enzymatic degradation). In addition studies were made of calf thymus DNA that had been degraded by means of the Waring blender. In all cases where the formaldehyde reaction was used, the effect of heat in enhancing the reaction was studied. Since dissolving in distilled water below pH 7.0 tends to disrupt the DNA double helix, resulting in the development of a considerable reaction with formaldehyde, the DNA was generally dissolved in 0.01 M NaCl prior to testing. The effect of distilled water at pH 7.0 and thereabouts was also studied in some cases as well as the effect of distilled water at a pH of about 8.5. In the latter case the double helix may be even more stable than it is in 0.01 M NaCl.

EXPERIMENTAL DETAILS

Cell nuclei were isolated from rat liver at pH 5.8 to 6.0 by the method described by Dounce ('55). These nuclei contained what we have termed *slightly degraded DNA*. The degradation results from the action of cytoplasmic DNA-ase which is

reased from cytoplasmic particles during homogenization. The method also was modified by holding the pH between 6.0 and 6.15 with the purpose of obtaining what we have termed *degraded* DNA. In this case the method for isolating the nuclei is more difficult to carry out and the yield is low, owing to the disruption of some of the nuclei than occurs when the pH is held between 5.8 and 6.0. DNA was isolated from the above two types of nuclei by the method of Kay et al. ('52).

DNA was also isolated from calf thymus by the same method (Kay et al., '52), and samples of this material were subjected to degradation by the action of a Waring blender. In these experiments 100 mg of DNA was dissolved in 200 ml of 0.01 M NaCl and then was allowed to remain for three minutes in a Waring blender running at full speed. After this the DNA was precipitated by adjusting the NaCl concentration to 1 M and adding one volume of 75% ethanol. The reprecipitated material was washed twice with 75% ethanol, twice with 95% ethanol, and finally was dried with ethyl ether.

DNA-ase I-degraded DNA was obtained by treating a 100 mg sample of calf thymus DNA dissolved in 100 ml of 0.01 N NaCl solution with 2.5 mg crystalline DNA-ase I (Worthington) for 5 minutes in the presence of 0.002 M Mg-acetate. The degraded DNA was isolated from the solution by adding an equal volume of 95% alcohol in the presence of 1 M NaCl. The precipitated DNA was washed twice with 75% alcohol, and twice with 95% alcohol, and finally was dried with ether.

The formaldehyde reaction was carried out according to the directions of Fraenkel-Conrat ('54).

RESULTS AND DISCUSSION

We have found that the only DNA samples showing any evidence of the presence of single strand material before heating, as judged by the formaldehyde reaction, were the *degraded* DNA samples and the calf thymus DNA degraded by means of the Waring blender. This finding correlates with the primer activities of the samples, which were measured by Dr. F. J. Bollum,² since only the *degraded* liver DNA (degraded by DNA-ase *in situ*) and the Waring

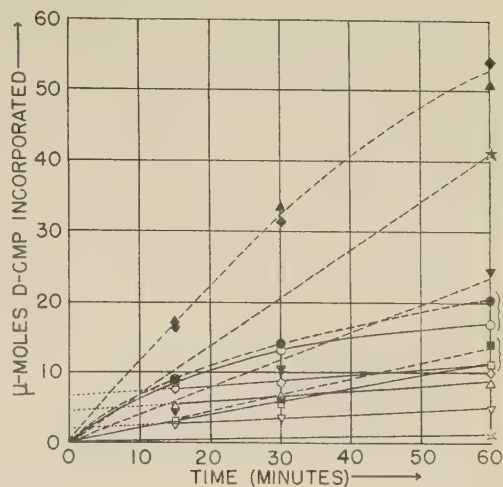


Fig. 1 Priming activities of various types of DNA described in text, before and after heating. ○, ●, *Degraded* DNA (from rat liver nuclei) after treatment with 6 M urea. □, ■, *Waring blender-degraded* DNA. △, ▲, *Degraded* DNA (from rat liver nuclei). ◇, ◆, *Partially degraded* DNA (from rat liver nuclei). ∇, ▼, DNA extracted from rat liver nuclei isolated at pH 4.0. ×, ★, Calf thymus DNA. Solid line with open symbols indicates priming activity before heating and dotted line with solid symbols refer to values obtained after heating. All heated samples were heated at 100°C for 10 minutes. Data for all curves obtained from equal aliquots of the same DNA solution. (This is also true for all subsequent figures.)

blender-degraded calf thymus DNA showed appreciable primer activity before heating (see fig. 1).

We have subjected all DNA samples to analysis of base ratios according to the method of Wyatt ('55), as well as to the formaldehyde reaction before and after heating. The results of analyses for base ratios are shown in table 1. The base ratios of the enzyme-degraded samples (*slightly degraded* and *degraded*) are not normal for intact DNA, and this taken by itself indicates a certain tendency towards single strand formation. However it was necessary to treat the DNA samples with 0.3 N KOH at 37°C for 18 hours before carrying out determinations of base ratios, since all DNA samples isolated from pH 4 liver cell nuclei were contaminated with RNA to

² We are greatly indebted to Dr. F. J. Bollum of the Oak Ridge National Laboratories for carrying out a number of tests of DNA samples for primer activity in his system for DNA synthesis using thymus enzyme (Bollum, '59).

TABLE 1

DNA sample ¹	A	T	G	C	A/T	G/C	PU/Py	A + T G + C	% RNA before alkali treatment
1. Calf thymus	1.00	1.04	0.75	0.75	0.95	1.00	0.97	1.35	3.8
2. Whole rat liver	1.00	1.04	0.73	0.61	0.96	1.19	1.04	1.51	8.2
3. pH 6.0 nuclei, partly degraded	1.00	1.34	0.87	0.78	0.74	1.11	0.88	1.41	42
4. pH 6.0 nuclei degraded	1.00	1.26	0.81	0.89	0.79	0.91	0.84	1.32	33
5. pH 6.0 nuclei partly degraded, urea-treated	1.00	0.95	0.71	0.59	1.05	1.20	1.11	1.48	21
6. pH 6.0 nuclei, degraded, urea-treated	1.00	0.93	0.76	0.60	1.06	1.25	1.14	1.32	31
7. pH 4.0 rat liver nuclei, DNA isolated at pH 6.0 soluble RNA first re- moved by ex- traction with 0.9% NaCl	1.00	1.04	0.73	0.65	0.96	1.11	1.02	1.46	31
8. Rat liver nuclei isolated at pH 4.0, DNA isolated from them at pH 6.0. Soluble RNA not re- moved first	1.00	0.79	0.56	0.61	1.25	0.92	1.11	1.51	48
9. Rat liver nuclei isolated at pH 4.0, DNA iso- lated from them at pH 7.0 sol- uble RNA not removed first	1.00	0.76	0.42	0.61	1.29	0.74	1.06	1.76	45

¹ All samples treated with 0.3 N KOH at 37°C for 18 hours before analysis—DNA recovered by precipitation with perchloric acid. Each figure in the table represents the average of duplicate analyses.

the extent of 30 to 40% (see table 1), and even the calf thymus DNA contained 3 to 4% of RNA.³ As the *slightly degraded* DNA did not show a reaction with formaldehyde and hence did not exist to an appreciable extent in the form of single strand material, it is likely that any single strand formation indicated by analysis of base ratios was brought about by the action of the alkali in dissolving away degraded single strand fragments from more intact complementary chains.

The extent of reaction of the *degraded* DNA samples with formaldehyde before and after adding urea is shown in figures

2, 3, and 4. Control experiments with undegraded calf thymus DNA are illustrated in figure 5. A tendency towards the formation of single strands is indicated for the *degraded* DNA by the results of the

³ We now know that this difficulty could have been largely avoided by precipitating the DNA in all cases with ½ volume of ethanol instead of with one volume from the saline detergent solution. However RNA does not have primer activity in Bollum's system and moreover treatment with alkali is a standard method to remove RNA before doing analysis for base ratios on DNA, so that as far as we can see the presence of RNA in the DNA samples does not affect our conclusions in any important manner.

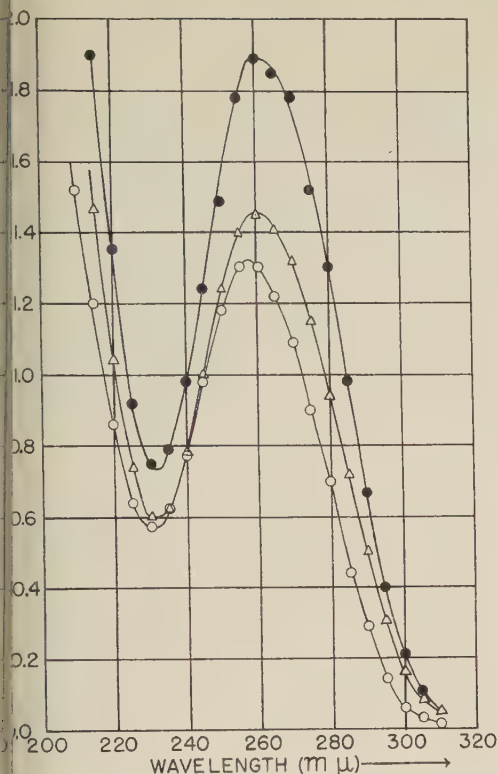


Fig. 2 U.V. spectrum of degraded DNA extracted from rat liver nuclei isolated at pH 6.0–6.5, dissolved in 0.01 M NaCl. ○, Degraded DNA, no formaldehyde, no heating. △, Degraded DNA after reaction with formaldehyde, no heating. ●, Degraded DNA, after heating at 100°C for 10 minutes, formaldehyde added after cooling. All samples dissolved in 0.01 M NaCl. Heating carried out for 10 minutes at 100°C. Concentration of formaldehyde where used approx. 1.3%. (Increase in formaldehyde concentration at controlled pH does not cause further change in spectrum.)

formaldehyde reaction, primer activity, and analysis of base ratios.

This urea-treated, degraded DNA is of considerable interest. It was originally hoped that urea treatment would cause the removal of small oligonucleotide single-strand fragments from their more intact complementary neighbor strands, as suggested above to explain the action of alkali on the degraded and slightly degraded samples. However the table of analysis of base ratios (Table 1) shows that this probably is not the case, and we believe from the results of the following experiments that the main action of the urea was to facilitate the re-

moval of contaminating RNA from the degraded DNA sample.

RNA isolated from yeast by Kirby's ('56) method was separated into two fractions by dissolving in 1 M NaCl and chilling to zero for several hours. A part of the material precipitated (the "less soluble fraction") and this was centrifuged off, leaving a supernatant solution from which a "soluble" RNA fraction was isolated by precipitation with two volumes of ethanol at 0.3°C.

Both the "less-soluble" and "soluble" yeast RNA fractions just described could be largely precipitated from 1 M NaCl solution at a concentration of 1 mg of RNA per ml if one volume of ethanol was added, but in the presence of 6 M urea the "soluble" fraction would not thus precipitate. Six molar urea did not however prevent the

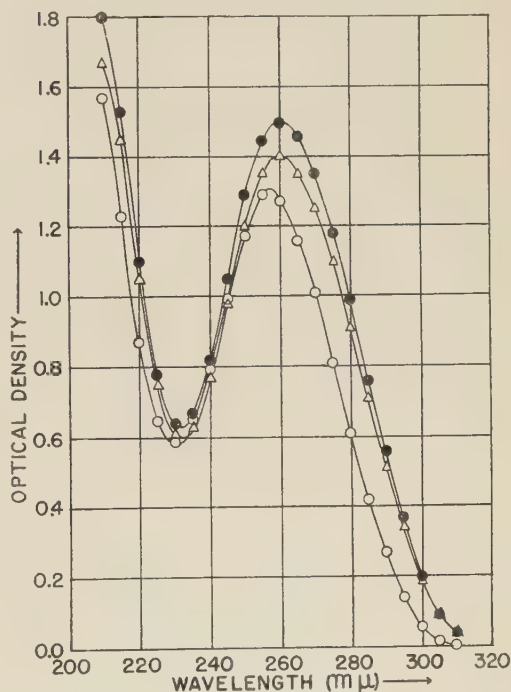


Fig. 3 U.V. spectrum of urea-treated degraded DNA extracted from rat liver nuclei isolated at pH 6.0–6.15. ○, Urea-treated degraded DNA. △, Urea-treated degraded DNA after reaction with formaldehyde, no heating. ●, Urea-treated degraded DNA after heating, formaldehyde added after cooling. All samples dissolved in 0.01 M NaCl. Heating carried out for 10 minutes at 100°C. Concentration of formaldehyde where used approx. 1.3%.

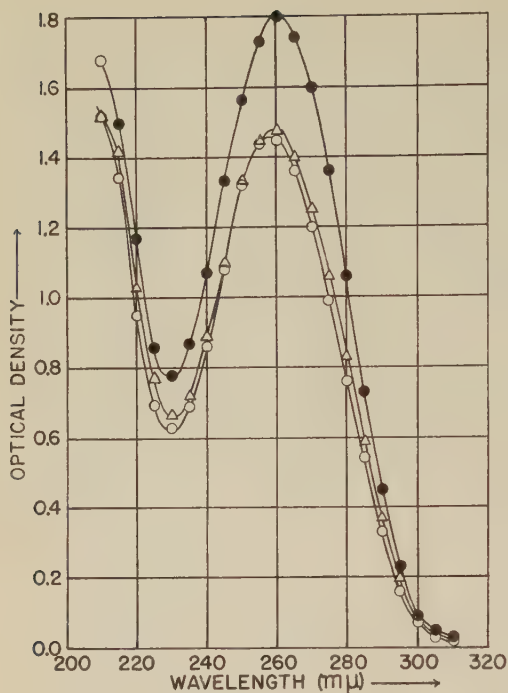


Fig. 4 U.V. spectrum of urea-treated degraded DNA extracted from rat liver nuclei isolated at pH 6.0–6.15. ○, Urea-treated, degraded DNA. △, Urea-treated degraded DNA after reaction with formaldehyde, no heating. ●, Urea-treated degraded DNA after heating, formaldehyde added after cooling. All samples dissolved in distilled water at pH 8.5; heated sample held at 100°C for 10 minutes. Concentration of formaldehyde where used approx. 1.3%.

precipitation of the "less-soluble" RNA fraction by one volume of alcohol under the same conditions used for the "soluble" fraction. It is thus clear that if much or most of the nuclear RNA is similar to the "soluble" fraction of yeast RNA isolated by Kirby's method, 6 M urea would be expected to aid in its removal from the DNA during the reisolation procedure. That this is actually the case was confirmed from the results of analysis for ratios of uracil to thymine which indicated that less RNA was present in the degraded urea-treated samples than in the degraded samples that had not been treated with urea.

It will be noted from table 1 that the results of analysis for base ratios of the degraded, urea-treated DNA show that after the urea treatment, the base ratios tend to approach the values for undergraded DNA,

although they still are not normal. We attribute this finding to the removal by the treatment with urea of the most degraded fragments of the DNA, leaving less degraded DNA which is more nearly normal in base ratios.

It should be noted that urea does not produce any effect on the *slightly degraded* DNA except to shift the base ratios somewhat towards those for undergraded DNA, nor does it have any effect at all, as far as is known, on intact calf thymus DNA. It may be that in the case of the *less-degraded* DNA, the contaminating RNA co-precipitates more extensively with the DNA in the presence of 6 M urea, because the DNA is still somewhat fibrous.

In regard to the action of DNA-ase I on purified calf thymus DNA in the presence of 0.002 M MgAc₂, it was found that only about 20% as much enzyme was required to produce a given amount of degradation as is required to degrade DNA-nucleoprotein *in situ* (Dounce et al., '57). However, studies of the calf thymus DNA degraded by crystallized DNA-ase showed results that were practically indistinguishable from the results of studies of the degraded

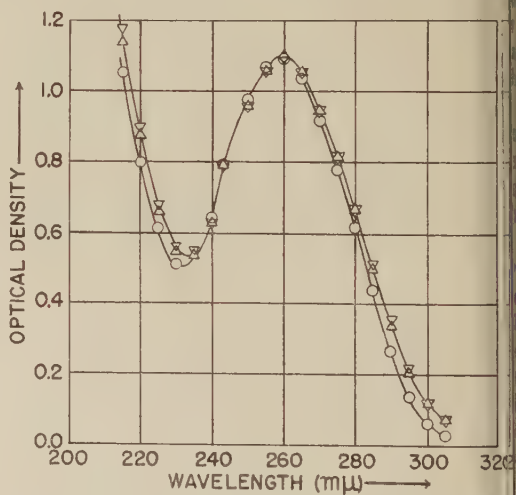


Fig. 5 U.V. spectrum of calf thymus DNA dissolved in 0.01 M NaCl under various conditions. ○, Calf thymus DNA. △, Calf thymus DNA after reaction with formaldehyde, no heating. ▽, Calf thymus DNA in 0.01 M NaCl, pH adjusted to 5.0, after reaction with formaldehyde, no heating. ●, Calf thymus DNA in 0.01 M NaCl, pH adjusted to 5.0, after reaction with formaldehyde, no heating. All samples dissolved in 0.01 M NaCl in all cases. Concentration of formaldehyde where used approx. 1.3%.

from pH 6 liver cell nuclei, the only difference being that the formaldehyde reaction was slightly greater in the case of degraded DNA isolated from the nuclei. These experiments show that DNA-ase I has a certain tendency to attack each end of the DNA double helix to a different extent, even when purified DNA instead of nucleoprotein is used as substrate, owing to the production of a certain amount of single strand material after treatment with alkali; but the presence of one and residual protein seems to exert a non-specific hindrance to DNA-ase action rather than a specific effect on one end only of the double helix.

The Waring blender-treated material is of considerable interest. This has recently been converted in part to a degraded product in which there has been appreciable hydrogen bond rupture; but addition scission of the double helix to even shorter segments also must have occurred. This is indicated by the fact that at pH 8.5, the Waring blender-degraded samples do not show a reaction with formaldehyde, indicating a reversal of the denaturation process, but in spite of this original fibrous nature of the material is not restored when the DNA is precipitated at pH 8.5 by one volume of alcohol in presence of 1.0 M NaCl.

The effect of formaldehyde on the Waring blender-degraded DNA is shown in figure 6. The results clearly indicate a mixture of double-strand and single-strand material.

It is of interest that heat treatment has no apparent effect on the Waring blender-degraded calf thymus DNA, than on native DNA. We interpret this to indicate that the heat denaturation, which certainly must be, is more reversible in the cases of the degraded samples, at least as judged by the formaldehyde reaction, than is the heat denaturation of native DNA.

One other point that requires comment is the difference in behavior towards heat of the degraded DNA (degraded by DNA-ase *in situ*) and the Waring blender degraded-DNA (see figs. 2 and 6). It is possible that the increase in primer activity (1) and in reactivity towards formaldehyde of the degraded DNA may be caused by the presence of RNA in the latter which

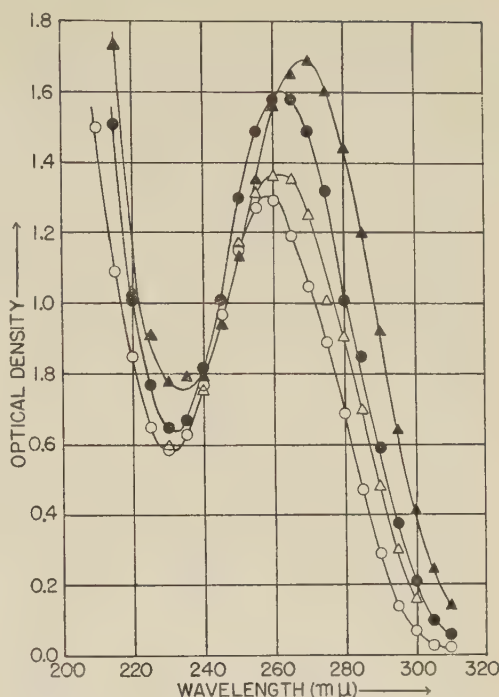


Fig. 6 U.V. spectrum of Waring blender-degraded DNA in 0.01 M NaCl, after reaction with formaldehyde under various conditions. ○, Waring blender-degraded DNA. Δ, Waring blender-degraded DNA after reaction with formaldehyde, no heating. ●, Waring blender-degraded DNA after heating for 10 minutes at 100°C, formaldehyde added after cooling. ▲, Waring blender-degraded DNA after heating, formaldehyde added before cooling solution. Heat-treated sample held at 100°C for 10 minutes. All samples dissolved in 0.01 M NaCl. Concentration of formaldehyde where used approx. 1.3%.

prevents reversal of the heat denaturation on cooling.

GENERAL DISCUSSION

It appears that the action of DNA-ase I on DNA does have a certain tendency to produce more cleavage in one of the two strands of the DNA double helix than in the other. However little difference was noted between the action of the DNA-ase on purified calf thymus DNA and on DNA-nucleoprotein of rat liver *in situ*, and hence the hypothesis that histone might sterically hinder the action of DNA-ase I on one of the two strands of the double helix, does not seem to be valid. Moreover we have not been able to show that single strand cleavage from DNA-ase I action occurs without

concomitant double strand scission. Hence the hypothesis that single strand formation might be brought about by DNA-ase I action prior to DNA replication is not favored by our results. It is true that we have not been able to work under completely physiological conditions but deviations from the latter would not seem to invalidate these conclusions, as far as we can see at the present time.

At least one other enzyme that might conceivably cause single strand formation from double strand DNA is phosphodiesterase, which is an exo-enzyme. However Bollum found no DNA synthesis in his system using thymus enzyme in the presence of phosphodiesterase with double strand DNA (Bollum, '60), although the addition of DNA-ase I to the same system in place of the phosphodiesterase did permit DNA synthesis to occur.

We believe that the reason for the latter observation is explained by the work reported in this paper. Our results would seem to indicate that the DNA synthesis which occurs *in vitro* after action of DNA-ase on the primer DNA probably does not represent a process that takes place under physiological conditions *in vivo*.

SUMMARY

1. The action of DNA-ase I on the nucleoprotein complex of the cell nucleus was investigated by allowing DNA-ase I of rat liver, liberated from cytoplasmic particles during homogenization of the liver, to act on cell nuclei present in the homogenate. A *degraded* and a *partially degraded* DNA was isolated from nuclei which had been isolated from the homogenates at pH 6.0 to 6.15 and from 5.8 to 6.0 respectively in the presence of very dilute citric acid.

2. The *degraded* DNA showed some activity in Bollum's system for DNA synthesis by thymus enzyme, and also showed some reactivity with formaldehyde without being heated, and on analysis showed abnormal base ratios. These results indicate a tendency toward single strand production, although concomitant double strand scission also occurs. The same was true for DNA degraded by action of a Waring blender.

3. Purified calf thymus DNA treated with crystallized DNA-ase I behaved simi-

larly to the degraded DNA isolated from the rat liver nuclei. Hence it can be concluded that the action of DNA-ase on DNA *in situ* during isolation of the nuclei is not affected qualitatively in an important way by the presence of protein in the nuclei (histone or residual protein).

Quantitatively, the reaction appears to be considerably slower in the presence of these proteins.

4. Treatment of the degraded DNA from the nuclei with 6 M urea did not appear to increase the amount of single-strand material present but probably facilitates the removal of contaminating RNA.

5. It is concluded from the results given above that formation of single-strand DNA by an attack of DNA-ase I on one of the two strands of double helix DNA prior to DNA replication *in vivo* probably does not occur.

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Crystalline Luciferin from Live *Cypridina*^{1,2}

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The small ostracod crustacean, *Cypridina*, produces a brilliantly luminescent secretion wherein light emission accompanies the aerobic oxidation of a substrate, luciferin, catalyzed by an enzyme, luciferase. Crystallization of the luciferin, extracted from whole, dried organisms, was achieved in 1957 by Shimomura, Goto and Iwata, partly with the aid of purification procedures developed by Harvey and his associates through many years of research in this system, (cf. Harvey, '52; Tsuji, Iwase and Harvey, '55).

A new approach in the efforts to isolate pure luciferin, involving the use of living organisms rather than dried organisms, was independently undertaken in 1957 by Johnson and Sie, with the cooperation of Haneda, Takatsuki, Sugiyama and Masuda in Shimoda. This approach was based on the premise that the luminescent secretion obtained from living organisms by electrical stimulation or other means should be relatively free of many diffusible substances contained in whole organisms, especially after degradation reactions that no doubt occur during the process of slow drying, usually accomplished by exposure to the sun. With living organisms, paper chromatograms of the luminescent secretion, drawn into methyl alcohol in a suction flask immediately after stimulation, tended to justify the original premise, but various circumstances made it difficult to obtain an amount of secretion sufficient for the purposes in view. On the other hand, extracts of initially living, whole organisms appeared to offer certain advantages over those of dried, whole organisms, e.g., in having a smaller number of impurities and a larger content of luciferin.

The present report is based on the further pursuit of this approach, which has led to a considerably simplified procedure for the isolation of luciferin and to its crystallization in appreciable quantities for the first time from summer organisms, whose luciferin has been found to be less in amount and more difficult to purify than that of organisms collected during the colder seasons of the year.

EXPERIMENTAL METHODS AND RESULTS

Warm methanol extracts

Anderson ('35) noted that, although the solubility of luciferin in different solvents varied widely with biochemical treatment of the material (starting with dried organisms), the active substance was spectacularly soluble in methanol under all conditions. A number of methanol extracts were prepared in 1957 at Shimoda, by plunging the living organisms into a large volume of methanol in a suction flask at room temperature, evacuating the air, shaking vigorously, sometimes in a warm water bath, then filtering through paper in a Buchner. The filtrate was always rich in yellow or greenish yellow substances, and highly fluorescent with a somewhat similar color in ultraviolet. Repeated extraction increased the yield of such substances, indicative of luciferin, and highly

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² Contribution number 121 from the Shimoda Marine Biological Station.

active in light emission with luciferase. These filtrates were subjected to various treatments before ultimately drying and sealing in vacuo for further fractionation in the Princeton laboratories. The light-emitting potency of the residues was found to vary with the previous treatment and with unknown factors, when tested later by means of a photomultiplier-light integrating apparatus. The best preparations had a potency of some 50,000 L.U. ("Light Units," arbitrarily defined units of total light emitted with luciferase) per milligram, whereas the ground powder of whole, dried organisms was often not more than 50 L.U./mg and did not exceed 300 L.U./mg in the most active of such material available.

The most nearly pure luciferin obtained from these preparations was by the following procedure. A net weight of 193 gm of living organisms was added to 600 ml methanol, the air evacuated, and the flask heated in a water bath at 65°C for one hour. The solution was cooled, filtered, and the methanol distilled off in vacuo, leaving about 70 ml of aqueous solution which was then extracted with benzene in a separatory funnel containing an argon atmosphere to retard the decomposition of the very oxygen-labile luciferin. The reddish benzene fraction, containing reddish solids at the interphase, exhibited a strong light-emitting activity with luciferase, but the yellowish aqueous fraction had much stronger activity. The latter was extracted with butanol, removing most of the luciferin, and the solvent was removed by vacuum distillation in a water bath at 80 to 90°C. The residue was taken up in methanol, transferred to an ampoule, the methanol then removed in vacuo, and the ampoule sealed.

Several weeks later the dry residue was found to have a light-emitting potency of 25,000 L.U./mg. It was then extracted with benzene in a Soxhlet, followed by dissolving in a small volume of Tsuji's ethanol-ethyl acetate-water solvent (Tsuji, '55) and placing on a cellulose column that had been deaerated by hydrogen purified over hot platinized asbestos. The column was developed in a hydrogen atmosphere by Tsuji's solvent that had been deaerated with hydrogen. Although 5

bands were visibly evident on an essentially similar column of material from dried organisms, after purification by prolonged extraction with benzene and ether followed by two cycles of a modified Anderson's ('36) benzoylation procedure (Shimomura et al., '57) only 3 bands were evident in the present instance. They were, (1) an orange colored, non-fluorescent band at the front, followed by a yellow, yellow fluorescent (in ultraviolet) band, and finally a colorless, blue fluorescent band. As expected from previous observations (Tsuji, Chase and Harvey, '55; Shimomura et al., '57), luminescence activity was associated with the yellow-yellow fluorescent band, of which 4 separate fractions, F1-F4, were collected and dried in vacuo. The third fraction from the front, F3, was richest in luciferin with a luminescence potency of 300,000 L.U./mg; portions of this fraction were used for spectroscopic studies and for ascending paper chromatography. The quantity of material available seemed too small to justify an attempt at crystallization.

Absorption spectrum, and changes on standing

The absorption spectrum of fraction F3 just after dissolving in methanol is illustrated in figure 1, curve A. It is very similar to that of the methanolic solution of crystalline luciferin reported by Shimomura et al., ('57, fig. 2, p. 930), differing only in having a shoulder rather than distinct maximum at 315 m μ , (a difference possibly due to a difference in pH; cf. discussion of fig. 3), and in having about a 50% larger ratio between optical density at 270 and the O.D. at 435 m μ , i.e., a ratio of approximately 3:1 instead of 2:1, indicative of some impurities that absorb in the ultraviolet.

The changes in absorption on standing aerobically at room temperature (fig. 1, curves B and C) are also similar to those of crystallized luciferin referred to above, and the isosbestic point at 390 m μ shows that no intermediary product accumulates. The changes in optical density of the 435 m μ maximum from the initial value, after two days, and after 8 days, closely parallel the changes in luminescence potency.

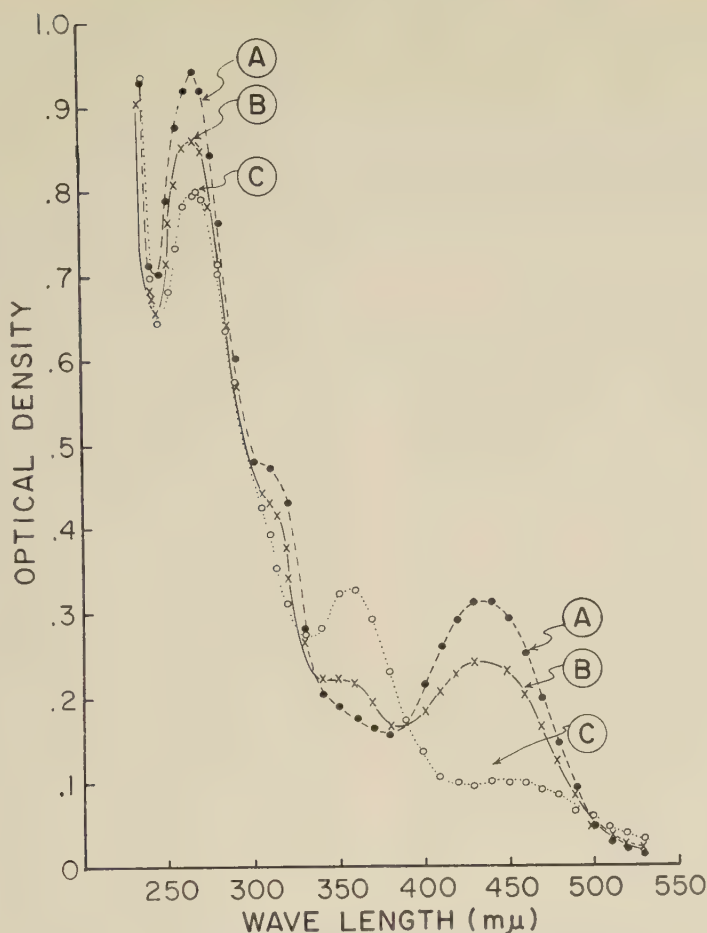


Fig. 1 Changes in the absorption spectrum of luciferin fraction F3, in a concentration of 0.108 mg/ml methanol, at room temperature in the presence of air, as described in the text. Curve A, just after dissolving; curve B, after two days; curve C, after 8 days.

is, the ratio of O.D.'s at these three successive times is 0.320:0.245:0.099, or 10:0.78:0.31, while the ratio of luminescence potency at the corresponding times was 300,000:234,000:45,000, or 10:0.78:0.15. These data leave little doubt that the absorption maximum at 435 mμ is characteristic of the substance, i.e., luciferin, that is active in light emission with luciferase.

Quantitative determinations of the total light potency of solutions of crystalline luciferin obtained in the present study have yielded values of 3.7 to 4.6 million L.U. per mg, varying with the luciferase preparation employed. Taking 4×10^6 L.U./mg as a mean value, the 300,000

L.U./mg in fraction F3 would indicate the presence of 7.5% luciferin, or 0.0081 mg/cm³ in the specimen involved. The initial optical density of 0.32 at 435 mμ would indicate 0.0171 mg/cm³, but when 85% of the activity has disappeared on standing, the O.D. remains 0.1, whereas the decomposition products of pure luciferin have practically no absorption at this wave length. Assuming that impurities absorbing at 435 mμ have an O.D. of 0.085, the amount of luciferin represented by the absorption at this point would be 0.0125 mg/cm³, or 11.5% luciferin in the specimen. Considering the various sources of error, these estimates of 7.5% luciferin, on the basis of light emitting potency, and

11.5% luciferin, on the basis of O.D. at 435 m μ , are in reasonably good agreement. Thus it should be possible to estimate with some assurance the actual quantity of pure luciferin in different lots of raw material through determinations of the number of light units emitted per mg, when tested with luciferase under appropriate conditions.

Paper chromatography

The results of ascending chromatography on Whatman paper no. 1 varied with the developing solvent, very likely due in part to the circumstance that facilities for complete anaerobiosis, essential for preventing oxidative decomposition of luciferin, were not available for the papers at that time, so developing was carried out at 2°C overnight. With Tsuji's solvent, a trace of orange-fluorescent substance at the front and a trace of blue fluorescent substance behind the broad, yellow colored, yellow fluorescent, active band was observed. With butanol two active bands, evidently corresponding to the alpha and beta luciferin of Mason and Davis ('52), were found at the origin and at an intermediate point from the front respectively; the latter band exhibited a blue fluorescence in UV, and though quite active in light emission with luciferase, was not in sufficient amount to exhibit color by ordinary light. With acetone most of the color, fluorescence and activity remained at the origin, but a trace of yellow color, yellowish fluorescence and luminescence activity with luciferase occurred at the front.

Chromatographic evidence for the existence of more than one species of luciferin molecules active in light emission with luciferase, such as the results of Mason and Davis ('52) and the results described above, is not fully convincing. Such evidence can be reasonably interpreted in terms of the influence of impurities, on the chromatographic behavior of one kind of luciferin. In unpublished experiments by Shimomura and associates, anaerobic paper chromatography of crystallized luciferin from dried organisms has revealed only a single yellow colored, yellow fluorescent, active band with Tsuji's solvent, and attempts to separate two distinctly different, pure luciferins by using

other solvents or combinations of solvents have been unsuccessful. Moreover, results of the present study indicate that the crystalline luciferin previously obtained from dried organisms is the same as that obtained more directly from freshly collected organisms, quickly frozen by ice and extracted at quite low temperatures, as described in the following paragraphs.

Cold methanol extracts

Experiments with extracts from *Shimoda* revealed that luciferin remained in solution in methanol at temperatures as low as -20°C, whereas considerable amounts of colorless impurities came out and could be removed by refrigerated centrifugation. In more recent experiments, live *Cypridina* collected at Tateyama were placed either in methyl alcohol pre-cooled to about -50°C in a polyethylene bottle, or were put into a thin polyethylene bag and then into a thermos jar containing dry ice. The latter method proved more convenient, and qualitative observations indicated that the organisms could be kept at the temperature of dry ice virtually indefinitely without perceptible loss of luminescence activity when thawed. By completing the initial extraction of luciferin without allowing the temperature ever to rise above -20°C, the extracts in effect were prepared from live organisms, and contained minimal amounts of fat and other impurities.

The yield of luciferin was augmented by several repeated extractions with two or three volumes of methanol per volume of organisms at -20 to -30°C in a vacuum flask with the air exhausted and with vigorous shaking. The cold solution was removed by rapid filtration through paper in a Buchner, with suction. A good yield could be obtained with a much smaller volume of cold methanol by homogenizing the organisms in a blender and filtering the brei with the aid of hyflo-supercel and washing with cold methanol. By this method it was more difficult to maintain the temperature below -20°C until filtration had been completed, and dry ice usually had to be added to the brei.

The initial fractionation was thus accomplished by the differential solubility of luciferin in methanol at low temperatures

The filtrate was always bright yellow in color, with strong yellow or greenish-yellow fluorescence in U.V. It was acidified by adding concentrated HCl to make the solution about 0.1 N acid, in order to convert the luciferin to a more stable state, and the methyl alcohol removed by vacuum distillation in a water bath at about 50°C, with a dry ice-acetone trap in the vacuum line, and a capillary stem supplied with pure H₂ at reduced pressure, in the flask to prevent bumping. The residual aqueous solution was extracted several times with benzene, whether or not colored substances went into the benzene. In some instances, reddish material containing active luciferin remained at the interface. Large, orange colored crystals of not fully pure luciferin were obtained from butanol solutions of such material by means of the usual further procedure described below.

Most of the luciferin in the acid, aqueous solution could be partitioned into butanol in a separatory funnel with an argon atmosphere. Saturated NaHCO₃ solution was added to neutralize the acid in the butanol layer, and the water layer was discarded. The butanol layer was concentrated to a small volume by vacuum distillation, and placed on a column of Brockman's standardized alumina in an atmosphere of hydrogen. The column assembly is diagrammed in figure 2.

On washing with butanol, the column usually showed three bands, in addition to some brownish, amorphous material, possibly denatured protein and inorganic salts, which remained on top: (1) a slowly moving, yellow colored, yellow fluorescent substance identified with active luciferin, (2) a pink to red material, inactive with luciferase moving more slowly than luciferin, and (3) a colorless, blue fluorescent material which stayed on the column. Mixtures of butanol and methanol, in proportions of 4:1 to 2:1 by volume, were used to elute the luciferin when it moved too slowly in butanol alone.

The eluate was dried by vacuum distillation in a water bath at 50 to 70°C, leaving a yellowish film, sometimes with small amounts of white material. The film dissolved readily in a few tenths of one ml of cold methanol. To the solution in a 15

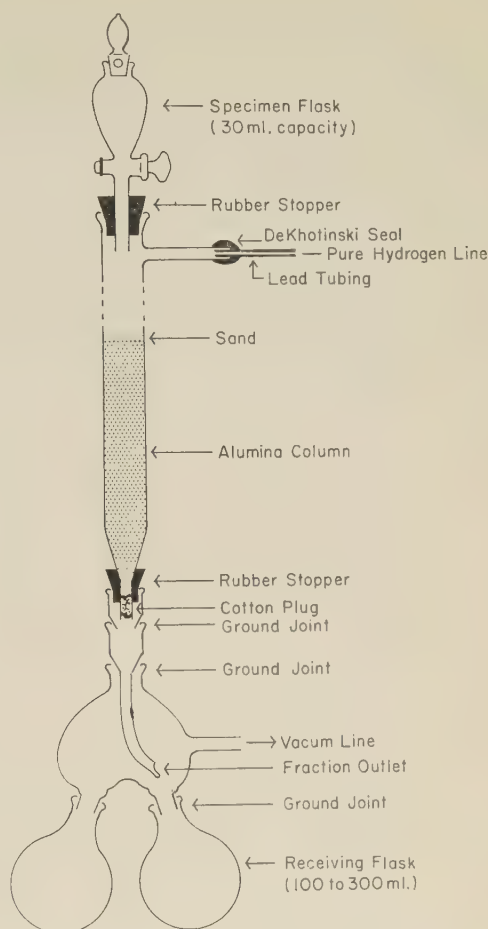


Fig. 2 Diagram of the apparatus used for anaerobic chromatography of luciferin on alumina. The column length was adjusted, from about 4 to 12 cm, in accordance with the estimated quantity of luciferin in the specimen involved. The whole assembly was evacuated and filled with hydrogen three successive times before introducing the specimen. The fraction outlet could be rotated into position over any one of three receiving flasks without exposing the eluate to oxygen.

ml centrifuge tube, three to 5 drops of distilled water were added, and a few small drops of 12 N HCl. In some instances, crystallization began almost at once. As a rule the centrifuge tube was placed in a vacuum desiccator over P₂O₅, with a beaker of KOH pellets on the porcelain support, the air was exhausted to 12 cm Hg, followed by filling with pure hydrogen and exhausting to this pressure

three times, and the desiccator placed in a refrigerator overnight. The crystals were then filtered off on a small sintered glass filter and dried in a vacuum desiccator. Figure 3 illustrates the absorption spectrum of the product, obtained in this manner, in comparison to that which was first reported (Shimomura et al., '57). The slight differences in the two curves are very likely due to slight differences in pH to which the absorption spectrum is particularly sensitive (Shimomura, '60).

By the above procedure, crystalline luciferin could be obtained in one day from

about 0.5 kg wet weight of organisms had been kept in dry ice. Purity and ease of crystallization were favored by speed in carrying out the procedure. The yield, however, was favored by starting with a larger amount of raw material, and 22 g wet weight, proved to be about optimum, though more than one day was required for purification. The partially purified luciferin, after partitioning into butanol, could be kept satisfactorily, for several days, if necessary, in a small flask in a thermos jar with dry ice.

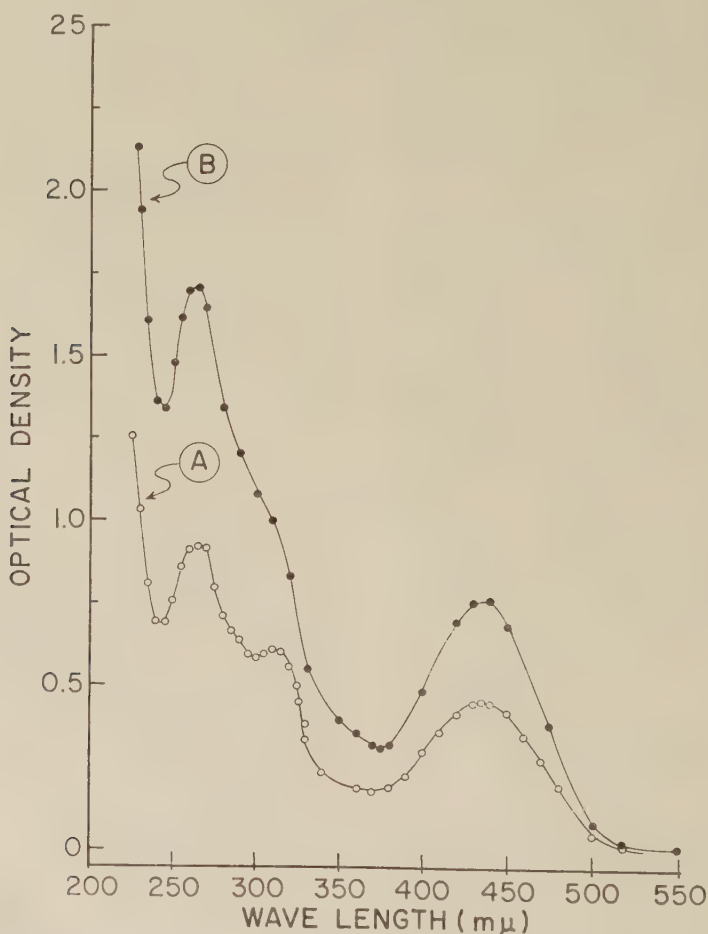


Fig. 3 Absorption spectrum of a methanolic solution of (A, open circles) crystalline luciferin 0.032 mg/cm³, from dried *Cypridina* (Shimomura, Goto and Hirata, '57, plotted from the original data) and (B, solid circles) of crystalline luciferin, 0.041 mg/cm³, obtained from cold methanol extracts of deep frozen, initially live *Cypridina*, as described in the text. Data for curve A were taken with a Beckman DU spectrophotometer, temperature 10°C. Data for curve B were taken on a Hitachi recording spectrophotometer, temperature 29°C.

The yield at best amounted to about 400 mg crystalline luciferin from 2 kg wet weight of frozen organisms. Assuming a 10% to 80% yield, the living organisms could contain an average of 0.0025 to 0.004% luciferin by weight. Since water constitutes about 75% of the wet weight, the actual yield would correspond to 80 mg per kg of dried organisms, provided no loss occurred during drying. The best yield, however, from organisms collected in cold seasons and dried, has been about 100 mg/kg dry weight, and heretofore it has been extremely difficult to obtain appreciable quantities of crystalline luciferin from summer *Cypridina*. It is reasonable to conclude that the luciferin content of the cold season organisms is at least an order of magnitude higher than that of summer organisms. If so, the cold methanol procedure should yield at best 400 mg crystalline luciferin per kg wet weight of winter organisms. Again assuming a 10% to 80% yield, the cold season, living organisms would contain between 500 and 1000 mg/kg, or between 0.050 and 0.0625% luciferin by weight. These figures are one or two orders of magnitude lower than the value of 1% to 10% used by Harvey (1933) in estimating that the minimum amount of luciferin needed for the production of visible light in reacting with luciferase was one part of luciferin in from 4 to 40 billion parts of water; an estimate of one part of luciferin in from 10^{10} to 10^{11} parts of water appears to be more nearly correct.

The light-emitting potency of the first crystalline luciferin (Shimomura et al., 1947) was estimated as being almost 40,000 times that of the starting material. Under favorable conditions of assay, pure luciferin would be expected to have a constant potency, whereas the actual content in the starting material is subject to wide variations, as discussed above. The increase in potency on purification, therefore, might be expected to range from perhaps 5,000 to 100,000 times depending on the activity of the starting material. Moreover, since the "potency" is subject to variation with the efficiency of light production, in both crude and fully purified systems, through the influence of numerous factors, such as pH, temperature, specific ions, etc., as dis-

cussed in detail elsewhere (Johnson, Eyring and Polissar, '54), the above range under appropriate conditions might well be even greater.

SUMMARY

A simplified method is described for purification of the substrate, luciferin, in the luminescent enzyme, luciferase, system of *Cypridina* by extraction of initially living, whole organisms with methanol.

Extraction in vacuo with hot (65°C) methanol yielded a product containing about 10% luciferin, by distilling off the methanol in vacuo, de-fatting the aqueous residue with benzene, partitioning into butanol, and chromatographing on a cellulose column with Tsuji's alcohol-acetate-water solvent.

Quantitative data revealed that the decrease in light-emitting potency of the product, on standing aerobically in methanolic solution two days at room temperature, was the same as the decrease in optical density at 435 m μ , viz., 22%. After 8 days, luminescence potency had decreased by 85%, with a 69% decrease in O.D. at this wavelength.

Extraction with cold (-20°C) methanol, using summer organisms kept at dry ice temperature since shortly after collecting, yielded crystalline luciferin in an amount of approximately 20 mg/kg wet weight, by the same purification procedure except that the chromatography was on a column of alumina with butanol as the solvent. The absorption spectrum of the product was essentially identical with that of the luciferin first crystallized from dried organisms.

The luciferin content of living *Cypridina* collected in summer is estimated as averaging 0.0025 to 0.004% by weight, and perhaps more than 10 times this amount in organisms collected in cold seasons.

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